



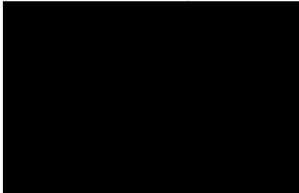
LARRY N. VANDERHOEF  
Chancellor at Davis

OFFICE OF THE VICE CHANCELLOR-ADMINISTRATION  
ONE SHIELDS AVENUE  
DAVIS, CALIFORNIA 95616-8540

JANET C. HAMILTON  
Vice Chancellor-Administration

May 11, 2000

Rae Newlands



*Dear C.E.P.E.  
please let me know what  
you glean from this - particularly  
my four.*

RE: California Public Records Act Request

*Best wishes and thank you  
for all that you are trying to do.*

Dear Mrs. Newlands,

This is in response to your February 25, 2000 letter in which you request all records pertaining to animals 24557, 30749, 23997, and 28545. We received your check in the amount of \$14.00 for the copying of the records and have enclosed a receipt.

The following records that are responsive to your request are enclosed:

*Love,  
x Rae x*

- 1) All of the pages from the health jackets of 24557, 30749, 23997, and 28545 (102 pages).
- 2) Animal Demographic/Medical Profiles for animals 24557, 30749, 23997, and 28545 (13 pages).
- 3) Protocols for Animal Use and Care that describe studies in which animals are involved - Protocol #8048 for animal # 30749; Protocol #8051 for animal #24557; Protocol # 8705 for animal #28545 (22 pages).
- 4) The California Regional Primate Research Center's (CRPRC) Standard Operating Procedure for feeding (3 pages).

We have redacted personally identifying information concerning individuals directly involved in research activities concerning primates due to verbal and physical harassment, including death threats, that have been made against these individuals. This information is withheld pursuant to section 6255 of the California Public Records Act which permits the University to not disclose records when the public interest served by not making the records public clearly outweighs the public interest served by disclosure of the record. In this case the public interest in withholding personally identifying information about these individuals due to actual harassment and threats of harassment that have occurred and continue to occur clearly outweighs the public interest in the disclosure of this information. See, e.g., Times Mirror Co. v. Superior Court, 53, Cal.3d 1325 (1991) (public interest in withholding the appointment calendars of the Governor of California due to "potential threat to the Governor's physical security" outweighed public interest in disclosure of the calendars); New York Times Co. v. Superior Court, 218 Cal.App.3d 1579 (1990) (names of persons who have violated water allocation limits may be withheld when there is evidence that release of such information may subject those persons to harassment or assault).

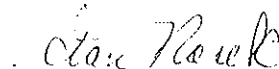
We have also redacted information that would identify the drug and its manufacturer as information that is subject to the California state law privileges for 'official information' (Evidence Code § 1040) and 'trade secret' (Evidence Code § 1060). 'Official information' subject to the privilege is information acquired in confidence by a University employee in the course of his or her duty and not open, or officially disclosed, to the public (Evidence Code § 1040). The pharmaceutical companies sponsoring the research trials have insisted that identifying information regarding the company and the drug name being studied be held in confidence by the University. There is a significant public interest in maintaining this confidence as release of such information would likely chill the interest of pharmaceutical companies in allowing the University to conduct the research trials, thereby foregoing the important research and teaching opportunities afforded to the University by such research trials.

The 'trade secret' privilege permits the owner of a trade secret to refuse to disclose the secret, and for the owner to prevent others from disclosing the secret. Information regarding the names of new drugs that were the subject of University studies falls within the definition of 'trade secret' as it is information that derives independent economic value from not being generally known to the public or to other persons who can obtain economic value from its disclosure or use and has been the subject of reasonable efforts to maintain its secrecy. The information that a particular drug is involved in a research study has economic value, both positive and negative, to the competitors of the drug manufacturer. It is for these reasons that the pharmaceutical companies have sought to ensure the secrecy of this information in their agreements with the University for conducting the trials.

In response to the questions you reiterated in your last letter, dated May 2, 2000, there are approximately 3,800 primates kept at the CRPRC. They currently have three species of primates: rhesus, cynomolgus, and titi monkeys.

Should you have any additional requests, please let me know.

Sincerely,



Stan Nosek  
Information Practices Coordinator  
(530) 752-6264

Enclosures

# PROTOCOL FOR ANIMAL USE AND CARE

Handwritten forms are not accepted



### Investigator

### Contact

APR 30 2001

Last Name: [REDACTED]  
 First: [REDACTED]  
 Middle: [REDACTED]  
 email: [REDACTED]  
 Department: Primate Center  
 Phone: [REDACTED]  
 Fax: [REDACTED]

Last Name: [REDACTED]  
 First: [REDACTED]  
 Middle: [REDACTED]  
 email: [REDACTED]  
 Department: Primate Center  
 Phone: [REDACTED]  
 Fax: [REDACTED]

Species (common names):	Number:	Source:
Macaca fascicularis	144	Primate Center

**Project Title** Effects of [REDACTED] on Ovariectomized Cynomolgus Monkeys

**Overnight housing location:** Primate Center Day use only : [REDACTED]

**Animals will be maintained by:**  Vivarium  Investigator (If Investigator maintained, attach husbandry SOP's.)

**Procedures:** Provide a one or two sentence layman's description of the procedures employed on the animals in this project. This information will help the animal care staff understand any conditions they may encounter while caring for your animals.

The animals will be given daily oral treatments of a drug used to alleviate the effects of osteoporosis. Test subjects will undergo periodic bone scans, biopsies, blood and urine sampling to monitor their progress.

**Special Husbandry Requirements:** Describe any special requirements your animals have with respect to food, water, temperature, humidity, light cycles, caging type, bedding, or any other conditions of husbandry.

Test animals will be fed a specially formulated monkey diet. All food will be removed from the animals' cages two hours prior to dosing and the animals will not be fed for one hour after dosing. Animals will be paired on a daily basis unless they are scheduled for procedures.

Other instructions for animal care staff: (check applicable entries)

#### Sick Animals

#### Dead Animals

#### Pest Control

- Call investigator
- Clinician to treat
- Terminate
- Necropsy
- Call Investigator
- Save for Investigator
- Bag for disposal
- Necropsy
- Call Investigator
- OK to use pesticides
- No Pesticides in animal area.

#### Hazardous Materials (only if in the animal room):

Infectious Agents?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No	Agent(s):	[REDACTED]
Radioisotopes?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No	Agent(s):	[REDACTED]
Chemical Carcinogens?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No	Agent(s):	[REDACTED]
Toxic Chemicals?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No	Agent(s):	[REDACTED]

Is the project already funded?  Yes  No  
Proposed Funding Source:  Extramural (private)

Previously approved?  Yes  No  
Previous protocol number (if any):

What Veterinarian or veterinary clinic will provide care for your animals? (check one)

Lab Animal Health Clinic ( 2-0514 )  
 VMTH Large Animal Field Service ( 2-0292 )

California Primate Research Center ( 2-0447 )  
 Another Veterinarian

If you checked "Another Veterinarian", please provide:

Veterinarian:  
Day phone:  
Emergency phone:

Address:  
Email:

*If your veterinarian is not affiliated with one of the three service units listed above, please contact the campus veterinarian, 2-2357 (email pctilman@ucdavis.edu) for current information about training and record keeping requirements.*

Summary of Procedures:

a) Briefly describe the overall Intent of the study. Include in your description a statement of your hypothesis, the objectives and significance of the study. Your target audience is a faculty member from a discipline unrelated to yours. Do not use jargon.

The overall intent of the study is to evaluate the effect of a proprietary drug [redacted] on ovariectomy-induced bone changes in cynomolgus macaques as a potential treatment for human post-menopausal osteoporosis. There will be five groups of 26 animals each. The control group without ovariectomy will be treated with vehicle only. A second control group will be ovariectomized and treated with vehicle. There will be three treatment groups that will be ovariectomized and treated once daily with [redacted] at 125 mg/kg, 250 mg/kg, and 625 mg/kg, respectively. All animals will be treated for two years during which time they will be monitored via blood and urine for biomarkers of bone turnover, Dual Energy X-Ray Absorptiometry (DXA) and Quantitative Computed Tomography (QCT) scans for measurements of bone mass, and bone biopsy for histomorphologic examination. All animals will be euthanized at the end of the study.

b) Procedures employed in this project:

Please check the appropriate boxes if any of these procedures will be employed in your project:

- Monoclonal Antibody Production \*\*       Food or water restriction       Special diets; food or water treatment.
- Polyclonal Antibody Production \*\*       Non-recovery surgical procedures       Induced illness, intoxication, or disease
- LD 50 or ID50 studies.       Survival surgical procedures       Death as an endpoint (see h below)
- catheters, blood collection, intubation       Multiple survival surgery       Trapping, banding or marking wild animals
- Prolonged restraint. (8 hrs+)
- Behavioral modification.
- Fasting prior to a procedure.       Aversive conditioning.

\*\* If this protocol only describes antibody production, you may use the attached antibody production page in lieu of completing section c below.

c) Describe the use of animals in your project in detail, with special reference to any of procedures checked above. Include any physical, chemical or biological agents that may be administered. List each study group, and describe all the specific procedures that will be performed on each animal in each study group. Use terminology that will be understood by individuals outside your field of expertise. (Note: This cell will expand to whatever length you require. You may make this section as long as you wish, but try to be concise. Some projects may require one or two pages.)

144 animals will undergo baseline radiography, bone mass determinations via dual energy X-ray absorptiometry (DXA) and quantitative computed tomography (QCT), blood sampling (approximately 7 ml per bleeding) and urine collection for bone biomarker analysis. DXA and QCT will be done under anesthesia and will take approximately one hour. 130 of these animals will be chosen for the study (the remainder will be returned to the colony) and will be randomly divided into five treatment groups. All animals will undergo fluorochrome labelling (IV administration of demeclocycline, 30 mg/kg) two times (14 and 7 days) prior to surgery. All animals will be anesthetized with isoflurane for iliac bone biopsy (approximately 8 mm<sup>2</sup>) and four groups will undergo bilateral ovariectomies (control group will have sham ovariectomy). One day post-op all animals will begin daily oral treatment with either vehicle (carboxymethylcellulose) or [REDACTED]. Treatment will last for 24 months. During this period the animals will be evaluated as follows:

- Day 1: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24hrs
- Week 4: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24hrs
- Week 26: DXA, QCT, Blood, Urine
- Week 39: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24hrs
- Week 52: DXA, QCT, Blood, Urine
- Week 54: Iliac Biopsy
- Week 74: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24hrs
- Week 78: DXA, QCT, Blood, Urine
- Week 91: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24 hrs
- Week 102: DXA, QCT, Blood, Urine
- Week 103: Pharmacokinetics - 1ml drawn at 0, 1, 2, 3, 6, 8, 16, 24hrs
- Week 104: Necropsy

d) Study Groups and Numbers: Define, in the form of a table, the numbers of animals to be used in each experimental group described above. The table may be presented on a separate page as an attachment to this protocol if you prefer. The Normal format should be three columns: Study Group, Procedure, Number of animals. The number of rows should follow from the number of study groups; you may add as many rows as you require. The chart must fully account for the number of animals you intend to use under this protocol. Assign each group to an invasiveness category according to the chart below.

Group	Procedures / Drugs	Number of Animals	Category
SHAM	Sham ovariectomy, iliac biopsies, blood samples, bone scans	26	3
OVX	Ovariectomy, iliac biopsies, blood samples, bone scans	26	3
S1	Ovariectomy, iliac biopsies, blood samples, bone scans, [REDACTED]	26	3
S2	Ovariectomy, iliac biopsies, blood samples, bone scans, [REDACTED]	26	3
S3	Ovariectomy, iliac biopsies, blood samples, bone scans, [REDACTED]	26	3

h) Neuromuscular blocking agents can conceal inadequate anesthesia and therefore require special justification. If you are using a neuromuscular blocking agent, please complete the following:

Why do you need to use a neuromuscular blocking agent?

n/a

What physiologic parameters are monitored during the procedure to assess adequacy of anesthesia?

n/a

Under what circumstances will incremental doses of anesthetics-analgesics be administered?

n/a

i) Adverse effects:

Describe any potential adverse effects of the experiment on the animals (such as pain, discomfort; reduced growth, fever, anemia, neurological deficits; behavioral abnormalities or other clinical symptoms of acute or chronic distress or nutritional deficiency)

Temporary pain and discomfort resulting from surgery. Bone labelling substances may induce transient vomiting during the IV infusion, therefore the infusions will be done slowly to alleviate this effect.

How will the signs listed above be ameliorated or alleviated? If signs are not to be alleviated or ameliorated by means of post-operative analgesics or other means, explain why this is necessary.

Post-operative administration of oxymorphone (.15 mg/kg) t.i.d. x 2d

Note: if any unanticipated adverse effects not described above do occur during the course of the study, a complete description of those effects and the steps taken to mitigate them must be submitted to the committee as an amendment to this protocol.

Is death an endpoint in your experimental procedure?  Yes  No

(Note: "Death as an endpoint" refers to acute toxicity testing, assessment of virulence of pathogens, neutralization tests for toxins, and other studies in which animals are not euthanized, but die as a direct result of the experimental manipulation). If death is an endpoint, explain why it is not possible to euthanize the animals at an earlier point in the study. If you can euthanize the animals at an earlier point, describe the clinical signs which will dictate that an animal will be euthanized.

[Empty box for explanation of death as an endpoint]

j) Literature search for alternatives and unnecessary duplication:

This section is specifically required by Federal law. You are required to conduct a literature search to determine that either 1) there are no alternative methodologies by which to conduct this study, or 2) there are alternative methodologies, but these are not appropriate for your particular study. "Alternative methodologies" refers to reduction, replacement, and refinement (the three R's) of animal use, not just animal replacement. You must also show that the study is not unnecessarily duplicative of other studies

What was the date on which you conducted this search?

4/4/98

List the databases searched or other sources consulted (there should be more than one). Include the years covered by the search.

Database Name	Years Covered	Keywords / Search Strategy
Medline	1990-Present	Osteoporosis, Research and Models

What were your findings with respect to alternative methodologies?

There are few rodent models available. However, this study is being conducted to meet FDA requirements which state that the study be done in a large animal remodeling species.

Has this study been previously conducted?  Yes  No

If the study has been conducted previously, explain why it is scientifically necessary to replicate the experiment.

k) Disposition of animals: At what point in the study, if any, will the animals be euthanized?

Animals will be euthanized after 24 months of treatment.

l) Methods of euthanasia: Even if your study does not involve killing the animals, you should show a method that you would use in the event of unanticipated injury or illness. If anesthetic overdose is the method, show the agent, dose, and route.

Species	Method	Drug	Dose (mg/kg)	route
M. fascicularis	Overdose	Pentobarbital	To effect	IV

m) Surplus animals: What will you do with any animals not euthanized at the conclusion of the project?

There will be no surplus animals.

**Categories of Invasiveness**

8048

Category	Description
1	<p>Little or no discomfort or stress</p> <p>Examples: domestic flocks or herds being maintained in simulated or actual commercial production management systems; the short-term and skillful restraint of animals for purposes of observation or physical examination; blood sampling; injection of material in amounts that will not cause adverse reactions by the following routes: intravenous, subcutaneous, intramuscular, intraperitoneal, or oral.</p>
2	<p>Minor stress or pain of short duration</p> <p>Examples: cannulation or catheterization of blood vessels or body cavities under anesthesia; minor surgical procedures under anesthesia, such as biopsies or laparoscopy; short periods of restraint beyond that required for simple observation or examination but consistent with minimal distress</p>
3	<p>Moderate to severe distress</p> <p>Examples: major surgical procedures conducted under general anesthesia, with subsequent recovery; prolonged (several hours or more) periods of physical restraint; induction of behavioral stresses such as maternal deprivation</p>
4	<p>Severe pain near, at or above the pain tolerance threshold</p> <p>Examples: exposure to noxious stimuli or agents whose effects are unknown; exposure to drugs, chemicals, or infectious agents at levels that markedly impair physiological systems and which cause death, severe pain, or extreme distress; Surgical experiments which have a high degree of invasiveness.</p>

Further descriptions of these categories are included in the instructions following this document.

e) Rationale for species and numbers: How did you determine that the species choice was appropriate and the number of animals in the groups above was the minimum number necessary to achieve sound scientific results?

The ovariectomy-induced bone loss model in cynomolgus macaques is the best model currently available for human post-menopausal osteoporosis and this study is intended to fulfill the FDA requirement for a pre-clinical study of bone quality in a non-rodent remodeling study. Due to the complexity of this study, large group numbers are required to complete statistical analysis. For those variables assessed only once during the study, a two-sided T-test will be used. For those variables assessed repeatedly, a two way (group, time) analysis of variance with repeated measures on time will be used. Significant group effects in these analyses will be evaluated using a Newman-Keuls test.

f) Surgery: If the project involves survival surgery, where will the surgery be conducted?

Building: Primate Center Animal Wing Rooms: CW1310, CW1316

Who will be the surgeons? [REDACTED]

g) Anesthetics, Analgesics, Tranquillizers, Neuromuscular blocking agents:

Post procedural analgesics should be given whenever there is possibility of pain or discomfort that is more than slight or momentary. If postoperative analgesics are not to be given, justify the practice under part (i) below.

Provide the following information about any of these drugs that you intend to use in this project.

Species	Drug	Dose (mg/kg)	Route	When and how often will it be given?
M. fascicularis	Ketamine	10	IM	Surgery (1x), bone scan (6x)
M. fascicularis	Medetomidine	.07	IM	Bone scan (6x)
M. fascicularis	Atipamezole	.07	IM	Bone scan (6x)
M. fascicularis	Atropine	.04	IM	Surgery (1x), biopsy (2x)
M. fascicularis	Isoflurane	1-290	Inhalant	Surgery (1x), biopsy (2x)
M. fascicularis	Oxymorphone	.15	IM	Post surgery, t.i.d. x 2d



n) **Project Roster:** Please provide the names of all the individuals who will work with animals on this project. This page will not be made available to the public. Give either the University Employee ID # or a valid UC Davis email address so that we can document training and occupational health compliance for regulatory agencies. Include all investigators, student employees, post-doctoral researchers, staff research associates, post-graduate researchers and laboratory assistants who will actually work with the animals. You don't need to include the staff of the vivarium in which your animals will be housed.

The principal investigator is responsible for keeping this roster current. If any staff is added or subtracted from this project, you must amend the protocol by sending the campus veterinarian a memo describing any changes.

Last Name	First Name	Middle Name	UC ID Number or SSN	Email Address

#### Occupational Health Program:

Supervisors must enroll their employees in the campus Occupational Health Program if the workers are at increased risk of illness or injury (such as allergy, physical injury, or infectious disease) because of their work. Enroll workers by having them complete an "Animal Contact History Form", available from Employee Health Services (phone 752-2330). For further information, visit our web site at <http://clueless.ucdavis.edu/health/> or read the UC Davis Policy & Procedure Manual 290-25.

#### Training:

Supervisors are responsible for insuring that their employees are adequate trained, both in the specifics of their job and in the requirements of the Federal Animal Welfare Act. EH&S offers free, basic wet labs in laboratory animal handling and techniques, and lecture format classes in the requirements of the Animal Welfare Act. To schedule a class for your unit, contact EH&S at 2-2364. Autotutorials are also available on the world wide web at <http://clueless.ucdavis.edu/>.


8048

Assurances for the Humane Care and Use of Vertebrate Animals:

Principal Investigator's Statement:

I have read and agree to abide by the UC Davis Policy and Procedure Manual section 290-30 (Animal Use and Care). This project will be conducted in accordance with the ILAR Guide for the Care and Use of Laboratory Animals, and the UC Davis Animal Welfare Assurance on file with the US Public Health Service. (These documents are available from the Campus Veterinarian and at <http://ohs.ucdavis.edu/>). I will abide by all Federal, state and local laws and regulations dealing with the use of animals in research.


I will advise the Animal Use and Care Administrative Advisory Committee in writing of any significant changes in the procedures or personnel involved in this project.

	Director	7/1/98
Principal Investigator	Rank / Title	Date

Committee Use Only Below

** Conditions necessary for Committee Approval:	
Final Disposition of this protocol:	
<input checked="" type="checkbox"/> Approved	
<input type="checkbox"/> Not Approved	
<input type="checkbox"/> Withdrawn by Investigator	
Date of Action:	APR 30 1998

I verify that the Institutional Animal Care and Use Committee of the University of California, Davis, acted on this protocol as shown above.

	APR 30 1998
Campus Veterinarian	Date

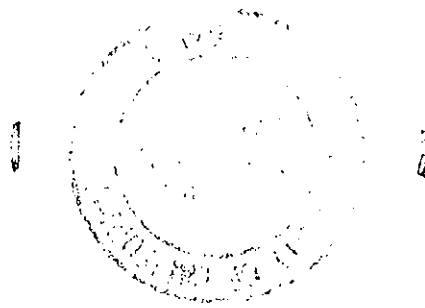
**CRPRC**

To: IACUC  
Re: Amendment to Protocol # 8048  
From: [REDACTED]  
Date: November 15, 1999

We would like to amend our protocol to include a second laparotomy in five animals. We previously ovariectomized 89 animals in this study. However, five of these have shown evidence of menses and elevated estradiol levels indicating retained or ectopic ovarian tissue. We would like to perform exploratory laparotomies on these animals to determine location of the ovarian tissue and to remove it. These surgeries will be performed by the CRPRC veterinary staff.

This amendment will not require any additional animals.

Possible adverse effects include those listed in the protocol such as post operative pain, suture dehiscence , or infection. Animals will be treated with oxymorphone (0.15 mg/kg TID for 2 days post operatively). Any other complications will be treated at the discretion of the CRPRC veterinary staff.



**The University of California at Davis**

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Animal Use and Care Administrative Advisory Committee  
Office of the Campus Veterinarian  
Davis, California 95616

May 1, 1998

National Institutes of Health  
Office for Protection from Research Risks  
6100 Executive Boulevard  
MSC 7507, Suite 3B01  
Rockville, MD 20892-7507

The following application was reviewed and approved by the Animal Use and Care Administrative Advisory Committee on 04/30/98. Active protocols are reviewed annually.

**PTF COLONY ANIMAL HUSBANDRY**

Principal Investigator: [REDACTED]  
Institution: The University of California at Davis

This institution is accredited by the American Association for the Accreditation of Laboratory Animal Care (AAALAC). This institution has an Animal Welfare Assurance on file with the Office for Protection from Research Risks. The Assurance number is A3433-01.

Animal Use and Care Administrative Advisory Committee is constituted in accordance with U. S. Public Health Service (PHS) Policy and includes a member of the public and a non-scientist.

Phil Tillman D.V.M.  
Philip C. Tillman  
Campus Veterinarian

CALIFORNIA REGIONAL PRIMATE RESEARCH CENTER  
 ANIMAL DEMOGRAPHIC/MEDICAL PROFILE, REPORT 315  
 MON, APR 10, 2000

ANIMAL ID	CURRENT LOCATION	DATE	WT (KG)	DEMOGRAPHIC ACTIVITY, CLINICAL OBSERVATION, OR MEDICAL EVENT
MCY 30749	BB 4004-40	MAR26-98	2.500	ACQUIRED TO QU4-4
		APR01-98	2.400	
		APR14-98	2.500	
		APR28-98	2.500	SERUM BANK SAMPLE
		MAY13-98	2.400	
		MAY27-98	2.500	
		JUN10-98	2.500	
		JUL02-98	2.490	MOVED FROM QU4-4 TO BB4004-39
		JUL09-98		IMMUNIZATION: CODE "K" IMMUNIZATION: TETANUS CLINICAL TREATMENT ADMINISTRATION OF DRUG OR SUBSTANCE, INTRAMUSCULAR IRON DEXTRAN 30 DAYS
		JUL16-98	2.300	
		JUL29-98	2.380	
		AUG12-98	2.370	
		AUG26-98	2.390	MOVED FROM BB4004-39 TO H01333-3
				CLINICAL TREATMENT ADMINISTRATION OF DRUG OR SUBSTANCE, INTRAMUSCULAR OXYMORPHONE 2 DAYS SURGERY BIOPSY, BY SURGICAL EXPOSURE ILIAC CREST EXCISION OVARY
		AUG27-98	2.390	MOVED FROM H01333-3 TO BB4004-39
		SEP09-98	2.450	
		SEP23-98	2.440	
		OCT04-98	2.440	
		OCT07-98	2.410	MOVED FROM BB4004-39 TO BB4004-58
		OCT22-98	2.410	
		NOV04-98	2.490	IMMUNIZATION: TETANUS
		NOV18-98	2.490	
		DEC02-98	2.490	
		DEC16-98	2.510	
		DEC30-98	2.520	
		JAN13-99	2.490	
		JAN27-99	2.540	

CALIFORNIA REGIONAL PRIMATE RESEARCH CENTER  
 ANIMAL DEMOGRAPHIC/MEDICAL PROFILE, REPORT 315  
 MON, APR 10, 2000

ANIMAL ID	CURRENT LOCATION	DATE	WT(KG)	DEMOGRAPHIC ACTIVITY, CLINICAL OBSERVATION, OR MEDICAL EVENT
XCY 30749	BB 4004-40	FEB10-99	2.490	
		FEB24-99	2.510	
		FEB25-99		
		MAR10-99	2.550	MOVED FROM BB4004-58 TO BB4004-40
		MAR24-99	2.530	
		APR07-99	2.520	
		APR21-99	2.530	
		MAY05-99	2.510	
		MAY19-99	2.500	
		JUN02-99	2.480	
		JUN16-99	2.490	
		JUN30-99	2.480	SERUM BANK SAMPLE
		JUL14-99	2.480	
		JUL28-99	2.490	
		AUG11-99	2.520	
		AUG25-99	2.460	
		AUG26-99		MOVED FROM BB4004-40 TO BB4004-58
		AUG31-99		MOVED FROM BB4004-58 TO H01602-3
		SEP01-99		CLINICAL TREATMENT ADMINISTRATION OF DRUG OR SUBSTANCE, INTRAMUSCULAR OXYMORPHONE 2 DAYS SURGERY BIOPSY
		SEP02-99		MOVED FROM H01602-3 TO BB4004-58
SEP08-99	2.430			
SEP22-99	2.440			
OCT06-99	2.440			
OCT20-99	2.390			
NOV03-99	2.480			
NOV17-99	2.440			
DEC01-99	2.450			
DEC15-99	2.510			
DEC29-99	2.540			
JAN12-00	2.480			
JAN26-00	2.480			
FEB09-00	2.440			
FEB23-00	2.500			
MAR02-00		MOVED FROM BB4004-58 TO BB4004-40		
MAR08-00	2.540			
MAR22-00	2.530			
APR05-00	2.570			

\*\*\* END ANIMAL XCY 30749



CALIFORNIA PRIMATE  
RESEARCH CENTER

I.D. 64P10 PROJECT CODE

mcg 30749  
ANIMAL I.D.

SURGERY  
POST-OPERATIVE  
RECORD

INVESTIGATOR [REDACTED] REQUESTOR [REDACTED]

9 1 99  
DATE OF SURGERY

ANIMAL DATA: 11002 -3  
ROOM CAGE

F SEX 8 YR 2 MO AGE 2.46 KG WEIGHT

WORKORDER # 3734 PROCEDURE: Diaphragm biopsy

IMMEDIATE POST-ANESTHETIC

TIME	SITTING UP	HEAT LAMP	EXTUBATION	TURNUED	OBSERVATION AND TREATMENTS	INIT.
9:45	✓	✓	✓	✓	recovery smooth	SR
11:10	✓				BAR	SR

SUBSEQUENT DAYS POST-OP  
All Entries Must Be Dated, Timed, and Initialed

DATE	TIME	APPETITE	HYD	STOOL	ATTITUDE	INCISION CONDITION	OBSERVATION AND TREATMENTS	INIT.
9/2/93	7:00 AM	F	G	N	BAR	intact	D/C to home cage	SR

SURGERY POST-OPERATIVE RECORD



CALIFORNIA PRIMATE RESEARCH CENTER  
**INTERVENTION / SURGERY**

ANIMAL SP ID# DATE OF EVENT  
 MO DAY YR  
 MAI 30749 9 / 1 / 99

PROCEDURE: Iliac Crest Biopsy

ROOM: 1602

REQUESTOR: [REDACTED]

CAGE: 3

INVESTIGATOR: [REDACTED]

PROJECT: GCPD

WT: KG 2.46

LINE	SNOMED CODES	CODED BY:	SNOMED TERMS (OPTIONAL)
01	T- 1234A P- 11400		
02	T- P-		
03	T- P-		

DESCRIPTION OF PROCEDURES PERFORMED

An approximate 4 cm curvilinear incision was made over the right ventral iliac crest to expose the tendonous attachment of the muscles to the crest. This attachment was then incised and the muscle dissected away from the iliac crest. One approximate 0.5 g bone biopsy was collected using a bone saw and making a single cut extending laterally from the corner of the previous biopsy site along the iliac crest. The incision was closed in two layers with absorbable suture.

ANESTHETICS, IV FLUIDS, CONCURRENT MEDICATION

		DOSE	UNITS	TOTAL	ROUTE
1	Ketamine				
2	Atropine				
3	NSAID				
4	LPS				

POSTOPERATIVE CARE AND CONDITION

TIME IN:

TIME OUT:

Oxycodone TID x 2 days

SURGEON:

ASSISTANT:

ANESTHETIST:

Animal's Record

Data Entry

Surgery

Requestor/Veterinarian

CALIFORNIA PRIMATE RESEARCH CENTER <b>INTERVENTION / SURGERY</b>	ANIMAL SP ID# MCY 30749	DATE OF EVENT MO DAY YR 8/26/1998
---	-------------------------------	---

PROCEDURE: [REDACTED]	ROOM:
REQUESTOR: [REDACTED]	CAGE:
INVESTIGATOR: [REDACTED]	PROJECT: WT: 2.37 KG

LINE	SNOMED CODES	CODED BY:	SNOMED TERMS (OPTIONAL)
01	T- 11339	P- 11420	
02	T- 87000	P- 11000	
03	T-	P-	

**DESCRIPTION OF PROCEDURES PERFORMED**

An approximate 4 cm ventral midline incision was made exposing the linea which was also incised. The uterus was identified and exteriorized. An ovary was identified and a clamp placed proximal to the ovary. The ovarian pedicle is ligated with absorbable suture and the ovary was removed. This procedure was repeated on the second ovary. The abdomen was closed in 2 or 3 layers with absorbable suture. An approximate 4 cm curvilinear incision was made over the right iliac crest to expose the tendonous attachment of the muscles to the crest. This attachment was then incised and the muscle dissected away from the iliac crest. Two approximate 1 cm<sup>2</sup> bone biopsies were collected using a bone saw and making two ventral and one longitudinal cut along the iliac crest. The incision was closed in two layers with absorbable suture.

ANESTHETICS, IV FLUIDS, CONCURRENT MEDICATION					
1		DOSE	UNITS	TOTAL	ROUTE
2					
3					
4					

POSTOPERATIVE CARE AND CONDITION	
TIME IN:	
TIME OUT:	

SURGEON: [REDACTED]	ASSISTANT: N/A	ANESTHETIST: [REDACTED]
---------------------	----------------	-------------------------

*Animal's Record*                     
 *Data Entry*                     
 *Surgery*                     
 *Requestor/Veterinarian*

4831

VIRAL PRECAUTION

# CALIFORNIA PRIMATE RESEARCH CENTER

GLP10  
PROJECT CODE

MCY 30749  
ANIMAL I.D.

## HEMATOLOGY

8/26/98  
DATE OF SAMPLE

I.D. [REDACTED]  
INVESTIGATOR [REDACTED] REQUESTOR [REDACTED]

ANIMAL DATA: 4004 - 39 HOME ROOM CAGE F 7 YR 2 MO 2.37 KG SEX AGE WEIGHT  
PROCEDURE IS:  DIAGNOSTIC AID \_\_\_\_\_ COLONY MANAGEMENT \_\_\_\_\_ EXPERIMENTAL

CLINICAL SIGNS / PROBLEMS:	PRIOR THERAPY <input type="checkbox"/> NO <input type="checkbox"/> YES
HOSPITALIZED NO <input type="checkbox"/> YES <input checked="" type="checkbox"/> 1333 - ROOM CAGE	<input type="checkbox"/> 2-COLOR FACS CD4 = / $\mu$ l <input type="checkbox"/> 3-COLOR FACS CD8 = / $\mu$ l CD4/CD8 RATIO =

BLEEDING CONDITIONS:  Squeezed - limb pulled  Caught on run  Fasted \_\_\_\_\_ hrs  Anesthetized  Other \_\_\_\_\_

COMPLETE BLOOD COUNT: ELECTRONIC CELL COUNT, SMEAR EVALUATION, PLASMA PROTEIN, FIBRINOGEN

<input type="checkbox"/> ELECTRONIC CELL COUNT		<input type="checkbox"/> SMEAR EVALUATION: TOTAL WBC 8.1 X 10 <sup>3</sup> / $\mu$ l		PLATELETS	
<input type="checkbox"/> CORRECTED WBC _____ X 10 <sup>3</sup> / $\mu$ l		DIFFERENTIAL		<input checked="" type="checkbox"/> ADEQUATE <input type="checkbox"/> DECREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> INCREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input checked="" type="checkbox"/> LARGE PLATELETS <input checked="" type="checkbox"/> CLUMPED	
WBC	8.1 X 10 <sup>3</sup> / $\mu$ l	8-26-98	7-60%	8-26-98	8-26-98
RBC	6.63 X 10 <sup>6</sup> / $\mu$ l	METAMYELOCYTES		ERYTHROCYTE MORPHOLOGY	
HEMOGLOBIN	12.8 gm/dl	BAND NEUTROPHILS		<input checked="" type="checkbox"/> ESSENTIALLY NORMAL <input type="checkbox"/> HYPOCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4 <input type="checkbox"/> POLYCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4 <input type="checkbox"/> LEPTOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4 <input type="checkbox"/> POIKILOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4 <input type="checkbox"/> ANISOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4 <input type="checkbox"/> ROULEAUX <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4	
HEMATOCRIT	41.8 %	SEG. NEUTROPHILS 56 4536			
MCV	63 fl	LYMPHOCYTES 31 2511			
MCH	19.3 pg	MONOCYTES 11 891			
MCHC	30.6 pg/fl	EOSINOPHILS 2 162			
PLATELETS	4.00 X 10 <sup>5</sup> / $\mu$ l	BASOPHILS			
<input type="checkbox"/> RETICULOCYTES	% _____ X 10 <sup>5</sup> / $\mu$ l	OTHER		<input type="checkbox"/> PARTIALLY CLOTTED SAMPLE <input type="checkbox"/> PREDILUTE	
<input type="checkbox"/> PCV (CENTRIFUGED)	%	NRBC/100 WBC			
<input type="checkbox"/> PLASMA PROTEIN	8.5 gm/dl	COMMENTS:			
PLASMA COLOR:		<input checked="" type="checkbox"/> NO ABNORMALITIES <input type="checkbox"/> HEMOLYZED <input type="checkbox"/> ICTERIC <input type="checkbox"/> LIPEMIC			
<input type="checkbox"/> FIBRINOGEN	200 mg/dl				

4-98  
1-98

REPORTED BY: [REDACTED] REPORT DATE: 8-26-98

# CLINICAL HEMATOLOGY

CALIFORNIA PRIMATE RESEARCH CENTER

MCL 30749  
ANIMAL I.D.

I.D. PROJECT CODE

INVESTIGATOR REQUESTOR

SURGERY POST-OPERATIVE RECORD

8, 26, 98  
DATE OF SURGERY

ANIMAL DATA: 4004 - 39  
ROOM CAGE

F 7 YR 2 MO 2.37 KG  
SEX AGE WEIGHT

WORKORDER # 6648 PROCEDURE: Ovariectomy / Bone Biopsy

IMMEDIATE POST-ANESTHETIC

TIME	SITTING UP	HEAT LAMP	EXTUBATION	TURNUED	OBSERVATION AND TREATMENTS	INIT.
10:45	yes	yes	yes	-	sitting when placed into cage - quiet	M
11:00	✓	✓	✓	-	sitting quietly	R
11:15	✓	✓	✓	-	sitting quietly	R
11:30	-	-	-	-	clearly can alert myself	R
12:00	✓	✓	✓	-	moderate	R
1 PM	✓	✓	-	-	animal B.P.D.	R
1:30 PM	✓	✓	-	-	looks good	R

SUBSEQUENT DAYS POST-OP  
All Entries Must Be Dated, Timed, and Initialed

DATE	TIME	APPETITE	HYD	STOOL	ATTITUDE	INCISION CONDITION	OBSERVATION AND TREATMENTS	INIT.

SURGERY POST-OPERATIVE RECORD

654 16410  
I.D. PROJECT CODE

CALIFORNIA PRIMATE  
RESEARCH CENTER

MC/ 30749  
ANIMAL I.D.

31675

URINALYSIS

INVESTIGATOR [REDACTED] REQUESTOR [REDACTED]

7/2/98  
DATE OF SAMPLE

ANIMAL DATA BB4004-39  
ROOM CAGE



F 7 YR 0 MO 2.49 KG  
SEX AGE WEIGHT

PROCEDURE IS DIAGNOSTIC AID COLONY MANAGEMENT  EXPERIMENTAL

SUSPECTED DIAGNOSIS	METHOD OF COLLECTION CYSTOCENTESIS
HOSPITALIZED <input type="checkbox"/> NO <input type="checkbox"/> YES	ROOM CAGE

long

Transparency	Hazy
Color	Yellow
Specific Gravity	1.006
Reaction pH	8.5
Protein	Ø
Glucose	Ø
Ketone	Ø
Bilirubin	Ø
Occult blood	Trace
CREATININE (TO IDEX, 7.2.98)	22.0 mg/dl
Miroscopic Sediment:	2.0 CC URINE
Casts	1+ (rare) granular
WBC	0-1/hpf
RBC	Ø
Epithelial cells	1+ (rare) renal
Crystals	Ø
Bacteria	Ø
Oil droplets	Ø
Sperm	Ø
Remarks:	marked amorphous matters

REPORTED BY: [REDACTED]

REPORT DATE: 7-2-98

CLINICAL URINALYSIS

3615  
 VIRAL PRECAUTION

# CALIFORNIA PRIMATE RESEARCH CENTER

6X54 / GLP10  
I.D. PROJECT CODE

MCY 30749  
ANIMAL I.D.

## HEMATOLOGY

INVESTIGATOR REQUESTOR

7/2/98  
DATE OF SAMPLE

ANIMAL DATA: BB4004-39  
HOME ROOM CAGE

F 7 YR 0 MO 2.49 KG  
SEX AGE WEIGHT

PROEDURE IS: \_\_\_\_\_ DIAGNOSTIC AID \_\_\_\_\_ COLONY MANAGEMENT  EXPERIMENTAL

CLINICAL SIGNS / PROBLEMS:	PRIOR THERAPY <input type="checkbox"/> NO <input type="checkbox"/> YES
HOSPITALIZED NO <input checked="" type="checkbox"/> YES <input type="checkbox"/>	<input type="checkbox"/> 2-COLOR FACS CD4 = / $\mu$ l
ROOM CAGE	<input type="checkbox"/> 3-COLOR FACS CD8 = / $\mu$ l
	CD4/CD8 RATIO =

BLEEDING CONDITIONS:  Squeezed - limb pulled  Caught on run  Fasted \_\_\_\_\_ hrs  Anesthetized  Other \_\_\_\_\_

COMPLETE BLOOD COUNT: ELECTRONIC CELL COUNT, SMEAR EVALUATION, PLASMA PROTEIN, FIBRINOGEN

<input type="checkbox"/> ELECTRONIC CELL COUNT			<input type="checkbox"/> SMEAR EVALUATION: TOTAL WBC 7.9 X 10 <sup>3</sup> / $\mu$ l			PLATELETS		
<input type="checkbox"/> CORRECTED WBC _____ X 10 <sup>3</sup> / $\mu$ l			DIFFERENTIAL			<input checked="" type="checkbox"/> ADEQUATE		
WBC	7.9	X 10 <sup>3</sup> / $\mu$ l	Ad 7-2-98 7.6-98-7W			<input type="checkbox"/> DECREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3		
REC	6.16	X 10 <sup>6</sup> / $\mu$ l	METAMYELOCYTES			<input type="checkbox"/> INCREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3		
HEMOGLOBIN	11.6	gm/dl	BAND NEUTROPHILS			<input type="checkbox"/> LARGE PLATELETS		
HEMATOCRIT	37.3	%	SEG. NEUTROPHILS 41 3239			<input checked="" type="checkbox"/> CLUMPED		
MCV	61	fl	LYMPHOCYTES 50 3950			ERYTHROCYTE MORPHOLOGY		
MCH	18.8	pg	MONOCYTES 9 711			<input checked="" type="checkbox"/> ESSENTIALLY NORMAL		
MCHC	31.1	pg/fl	EOSINOPHILS			<input type="checkbox"/> HYPOCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
PLATELETS	3.69	X 10 <sup>5</sup> / $\mu$ l	BASOPHILS			<input type="checkbox"/> POLYCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> RETICULOCYTES	%	X 10 <sup>5</sup> / $\mu$ l	OTHER			<input type="checkbox"/> LEPTOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> PCV (CENTRIFUGED)	%		NRBC/100 WBC			<input type="checkbox"/> POIKILOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> PLASMA PROTEIN	7.2	gm/dl	COMMENTS: <input type="checkbox"/> PARTIALLY CLOTTED SAMPLE <input type="checkbox"/> PREDILUTE			<input type="checkbox"/> ANISOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
PLASMA COLOR:								
<input checked="" type="checkbox"/> NO ABNORMALITIES								
<input type="checkbox"/> HEMOLYZED								
<input type="checkbox"/> ICTERIC								
<input type="checkbox"/> LUPEMIC								
<input type="checkbox"/> FIBRINOGEN	700	mg/dl						

REPORTED BY: [REDACTED]

REPORT DATE: 7/2/98

# CLINICAL HEMATOLOGY

White - Animal's Chart    Yellow - Laboratory    Pink - Requestor    Goldenrod - Clinical Pathologist

8794 CRXAL

CALIFORNIA PRIMATE RESEARCH CENTER

MCCY 30749

PROJECT CODE

ANIMAL I.D.

RADIOLOGY

7/2/98

DATE OF EXAM

INVESTIGATOR REQUESTOR



ANIMAL DATA: BB4004 - 39 HOMEROOM CAGE

SEX: F AGE: 7 YR 0 MO WEIGHT: 2.49 KG

HOSPITAL ROOM CAGE

PROCEDURE IS: \_\_\_\_\_ DIAGNOSTIC AID \_\_\_\_\_ COLONY MANAGEMENT \_\_\_\_\_ EXPERIMENTAL

TENT. DIAGNOSIS:

HISTORY:

QU Screen Out

EXAM REQUESTED

Head

- nasal cavity, teeth (upper/lower, R/L), mandible (R/L), maxilla (R/L), skull - routine

Neck

- cervical spine, soft tissues

Thorax

- routine, thoracic vertebra, esophagus, thoracic inlet

Abdomen

- routine, obstruction series, liver, intestinal tract, kidney, ureter bladder, uterus, prostate, lumbar vertebra, sacral vertebra, coccygeal vertebra, I.U., cystography, upper g.i., lower g.i., myelogram

Arm

- shoulder, R/L humerus, elbow joint, R/L radius-ulna, carpal joints, hand

Leg

- pelvis, R/L hip joint, femur, R/L knee joint, tibia-fibula, tarsal joints, foot

Ultrasound

Other: (Specify)

SPECIAL PROCEDURES:

Previous radiographs: Yes/No

Investigator:

Repeat studies required

at \_\_\_\_\_ days/weeks/months

Technique: Vertical, Table Top, Bucky

Film Type: Paraspred

Total No. Films: 2

Table with columns: cm, ma, time, kvp and rows: Lat., VD

RADIOGRAPHIC INTERPRETATION:

NAD

T-22100 / T-28800

CONCLUSIONS:

NORMAL

M-00100

REPORTED BY:

REPORT DATE: 7/2/98

**CALIFORNIA PRIMATE RESEARCH CENTER  
PHYSICAL EXAM AND EVALUATION/HEALTH CERTIFICATE**

SPECIES/ID: MACY 36749 LOCATION: BB-100434 DATE: 7/10/98  
 REASON FOR EXAM: ROUTINE PRE-SHIPMENT QU SCREEN EXPERIMENTAL  
 OTHER OUT

ORGAN SYSTEMS:			NAO=NO ABNORMALITIES OBSERVED	A=ABNORMAL	NE=NOT EXAMINED
1. INTEGUMENT	<u>NO</u>	A	NE	6. SPLEEN/L NODES	<u>NO</u> A NE
2. ORAL CAVITY	<u>NO</u>	A	NE	7. RESPIRATORY	<u>NO</u> A NE
3. EYES	<u>NO</u>	A	NE	8. DIGESTIVE	<u>NO</u> A NE
4. MUSCULOSKELET.	<u>NO</u>	A	NE	9. UROGENITAL	<u>NO</u> A NE
5. CIRCULATORY	<u>NO</u>	A	NE	10. OTHER	<u>NO</u> A <u>NE</u>

FEMORAL VESSELS: Right \_\_\_\_\_ Left \_\_\_\_\_  
 WEIGHT (kg) 2.49 DATE 7/2/98 CURRENT TB TEST 6/10/98

ABNORMAL FINDINGS:  
 \_\_\_\_\_  
 \_\_\_\_\_  
 \_\_\_\_\_  
 \_\_\_\_\_

**REPRODUCTIVE EVALUATION**

UTERUS: NAO A NE  
 ADHESIONS: MINOR MODERATE SEVERE  
 PREGNANCY STATUS:  
 PREGNANT: \_\_\_\_\_ NONPREGNANT: \_\_\_\_\_  
 GL (mm)= \_\_\_\_\_ UTERINE SIZE \_\_\_\_\_  
 BPD (mm)= \_\_\_\_\_ \_\_\_\_\_  
 FL (mm)= \_\_\_\_\_ CONTOUR/SHAPE \_\_\_\_\_  
 E/FHR (bpm)= \_\_\_\_\_ \_\_\_\_\_  
 Gest. Age (days) \_\_\_\_\_ \_\_\_\_\_  
 GENDER: M F

REPRODUCTIVELY SOUND \_\_\_\_\_ AREPRODUCTIVE \_\_\_\_\_ RE-EVALUATE \_\_\_\_\_ NOT EVALUATED \_\_\_\_\_

COMMENTS:  
 \_\_\_\_\_  
 \_\_\_\_\_

OVERALL CONDITION: EXCELLENT GOOD FAIR POOR

RECOMMENDATION: I CERTIFY TO THE BEST OF MY KNOWLEDGE THAT THIS ANIMAL HAS BEEN EXAMINED AND IS

SATISFACTORY FOR SHIPMENT COMMENT: \_\_\_\_\_

SATISFACTORY FOR PROJECT COMMENT: \_\_\_\_\_

OTHER Qu Screen Out COMMENT: \_\_\_\_\_

DATE: 7/6/98 EXAMINING VETERINARIAN: \_\_\_\_\_



301

# CALIFORNIA PRIMATE RESEARCH CENTER PARASITOLOGY

MCY 30749

ID: [REDACTED] PROJECT CODE: [REDACTED]  
INVESTIGATOR: [REDACTED] REQUESTOR: [REDACTED]

ANIMAL ID: [REDACTED]  
DATE OF SAMPLE: 4/28/98

ANIMAL DATA: QU4-4  
ROOM: CAGE:

SEX: F AGE: 6 YR 10 MO WEIGHT: 2.40 KG

PROCEDURE IS: \_\_\_\_\_ DIAGNOSTIC AID \_\_\_\_\_ COLONY MANAGEMENT  EXPERIMENTAL \_\_\_\_\_

CLINICAL SIGNS/PROBLEMS: <input type="checkbox"/> DIARRHEA  HOSPITALIZED NO <input type="checkbox"/> YES <input type="checkbox"/> ROOM: CAGE:	PRIOR THERAPY <input type="checkbox"/> NO <input type="checkbox"/> YES LIST ALL AGENTS:  SOURCE OF SPECIMEN: <input type="checkbox"/> FECES, Fresh catch <input type="checkbox"/> Composite <input checked="" type="checkbox"/> FECES, Cage sample <input type="checkbox"/> OTHER
--	--

PROCEDURE REQUESTED:

<input type="checkbox"/> DIRECT EXAMINATION	<input type="checkbox"/> SKIN SCRAPING EXAM
<input checked="" type="checkbox"/> CONCENTRATION	<input type="checkbox"/> STAIN FOR ACID FAST BACILLI
SEDIMENTATION <input checked="" type="checkbox"/>	<input type="checkbox"/> CRYPTOSPORIDIA / GIARDIA IFA
FORMALIN-ETHYLACETATE	
<input type="checkbox"/> FLOTATION	<input type="checkbox"/> OTHER
<input type="checkbox"/> ZINC SULFATE	

### FOR LABORATORY USE ONLY

APPEARANCE	CONSISTENCY: <i>formed</i>	COLOR: <i>brown</i>
EXAMINATION	<input type="checkbox"/> RBC: <i>Ø</i>	<input type="checkbox"/> WBC: <i>Ø</i>
	<input type="checkbox"/> OTHER:	

Balantidium coli	Entamoeba histolytica
Blastocystis hominis	Giardia lamblia
Chilomastix mesnili	Hexamita pitheci
Endolimax nana	<i>H</i> Iodamoeba butschlii
Entamoeba NOS	Trichomonas, NOS
Entamoeba coli	Trichuris trichiura
Entamoeba hartmanni	No Parasites Seen
Cryptosporidium IFA	Acid fast bacilli
Giardia IFA	Budding yeast

[REDACTED]

REPORTED BY: [REDACTED]

REPORT DATE: 6/5/98

# CLINICAL PARASITOLOGY

VIRAL PRECAUTION

# CALIFORNIA PRIMATE RESEARCH CENTER

## HEMATOLOGY

8794, CRX01  
I.D. PROJECT CODE

MCY 30749  
ANIMAL I.D.

[Redacted] INVESTIGATOR  
[Redacted] REQUESTOR

4/28/98  
DATE OF SAMPLE

ANIMAL DATA: QU4-4  
HOME ROOM CAGE

F 6 YR 10 MO 2.40 KG  
SEX AGE WEIGHT

PROEDURE IS: \_\_\_\_\_ DIAGNOSTIC AID  COLONY MANAGEMENT \_\_\_\_\_ EXPERIMENTAL

CLINICAL SIGNS / PROBLEMS:	PRIOR THERAPY <input type="checkbox"/> NO <input type="checkbox"/> YES LIST ALL AGENTS:
HOSPITALIZED NO <input type="checkbox"/> YES <input type="checkbox"/>	ROOM _____ CAGE _____

BLEEDING CONDITIONS:  Squeezed - limb pulled  Caught on run  Fasted \_\_\_\_\_ hrs  Anesthetized  Other \_\_\_\_\_

COMPLETE BLOOD COUNT: ELECTRONIC CELL COUNT, SMEAR EVALUATION, PLASMA PROTEIN, FIBRINOGEN

<input type="checkbox"/> ELECTRONIC CELL COUNT			<input type="checkbox"/> SMEAR EVALUATION: TOTAL WBC $\times 10^3/\mu\text{l}$ <input type="checkbox"/> CORRECTED WBC <i>available</i> $\times 10^3/\mu\text{l}$			PLATELETS		
WBC	6.4	$\times 10^3 / \mu\text{l}$	DIFFERENTIAL	%	$\mu\text{l}$	<input type="checkbox"/> ADEQUATE		
REC	5.74	$\times 10^6 / \mu\text{l}$	METAMYELOCYTES			<input type="checkbox"/> DECREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3		
HEMOGLOBIN	10.4	gm/dl	BAND NEUTROPHILS			<input type="checkbox"/> INCREASED <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3		
HEMATOCRIT	34.7	%	SEG. NEUTROPHILS			<input type="checkbox"/> LARGE PLATELETS		
MCV	60	fl	LYMPHOCYTES			<input type="checkbox"/> CLUMPED		
MCH	18.1	pg	MONOCYTES			ERYTHROCYTE MORPHOLOGY		
MCHC	30.0	pg/fl	EOSINOPHILS			<input type="checkbox"/> ESSENTIALLY NORMAL		
PLATELETS	4.10	$\times 10^5 / \mu\text{l}$	BASOPHILS			<input type="checkbox"/> HYPOCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> RETICULOCYTES	%	$\times 10^5 / \mu\text{l}$	OTHER			<input type="checkbox"/> POLYCHROMASIA <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> PCV (CENTRIFUGED)	%		NRBC/100 WBC			<input type="checkbox"/> LEPTOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
<input type="checkbox"/> PLASMA PROTEIN	7.5	gm/dl	COMMENTS:	<input type="checkbox"/> PARTIALLY CLOTTED SAMPLE <input type="checkbox"/> PREDILUTE		<input type="checkbox"/> POIKILOCYTOSIS <input type="checkbox"/> +1 <input type="checkbox"/> +2 <input type="checkbox"/> +3 <input type="checkbox"/> +4		
PLASMA COLOR:								
<input checked="" type="checkbox"/> NO ABNORMALITIES								
<input type="checkbox"/> HEMOLYZED								
<input type="checkbox"/> ICTERIC								
<input type="checkbox"/> LIPEMIC								
<input type="checkbox"/> FIBRINOGEN	200	mg/dl						

REPORTED BY: [Redacted]

REPORT DATE: 4-28-98

PROJECT CODE

# CALIFORNIA PRIMATE RESEARCH CENTER

ANIMAL I.D. 909

## MICROBIOLOGY

DATE OF SAMPLE 4/28/98

INVESTIGATOR: \_\_\_\_\_ REQUESTOR: \_\_\_\_\_

ANIMAL DATA: QU4 - 4<sup>7</sup>  
HOME ROOM CAGE

SEX F AGE 6 YR 10 MO WEIGHT 2.40 KG

PROCEDURE IS: \_\_\_\_\_ DIAGNOSTIC AID  COLONY MANAGEMENT \_\_\_\_\_ EXPERIMENTAL

CLINICAL SIGNS / PROBLEMS:  
 DIARRHEA  
  
HOSPITALIZED NO  YES   
ROOM \_\_\_\_\_ CAGE \_\_\_\_\_

PRIOR THERAPY  NO  YES  
LIST ALL AGENTS:  
  
SOURCE OF SPECIMEN(S)  
RECTAL SWAB

CULTURES REQUESTED	NEGATIVE RESULT	
	NEGATIVE	NO GROWTH
<input checked="" type="checkbox"/> SALMONELLA, SHIGELLA, YERSINIA, AEROMONAS		<input checked="" type="checkbox"/>
<input type="checkbox"/> CAMPYLOBACTER		
<input type="checkbox"/> YERSINIA SUSPECT (EXTRA SWAB)		
<input type="checkbox"/> AEROBIC		
<input type="checkbox"/> ANAEROBIC		
<input type="checkbox"/> FUNGI		
<input type="checkbox"/> OTHER _____		

DIRECT MICROSCOPIC EXAMINATION

- ### ORGANISMS IDENTIFIED
- - 
  - 
  - 
  - 
  - 
  - 
  -

SENSITIVITY TO ANTIMICROBIAL AGENTS: KIRBY-BAUER

ORGANISM NUMBER	AMIKACIN (AK 30)	AMPICILLIN (AM 10)	AUGMENTIN (AKC 30)	CEFAZOLIN (CZ 30)	CEFTRIAXONE (CRO 30)	CHLORAMPHENICOL (C 30)	CLINDAMYCIN (CC 2)	DOXYCYCLINE (D 30)	ENROFLOXACIN (ENO 5)	GENTAMICIN (GA 10)	NEOMYCIN (N 30)	OXACILLIN (OX 1)	PENICILLIN (P 10)	SULFA/ TRIMETH (SXT 25)	VANCOMYCIN (VA 30)

COMMENTS:  
REPORTED BY: \_\_\_\_\_

REPORT DATE: 4/28/98

# CLINICAL MICROBIOLOGY

White - Animal's Chart      Yellow - Laboratory      Pink - Requestor      Goldenrod - Clinical Pathologist

**CALIFORNIA PRIMATE RESEARCH CENTER  
PHYSICAL EXAM AND EVALUATION/HEALTH CERTIFICATE**

SPECIES/ID# MOY 30749 LOCATION QUILHA DATE 4/8/98  
 REASON FOR EXAM: ROUTINE PRE-SHIPMENT QU SCREEN EXPERIMENTAL  
 OTHER

ORGAN SYSTEMS: NAO=NO ABNORMALITIES OBSERVED A=ABNORMAL NE=NOT EXAMINED		
1. INTEGUMENT	<u>NAO</u> A NE	6. SPLEEN/L. NODES <u>NAO</u> A NE
2. ORAL CAVITY	<u>NAO</u> <u>(A)</u> NE	7. RESPIRATORY <u>NAO</u> A NE
3. EYES	<u>NAO</u> A NE	8. DIGESTIVE <u>NAO</u> A NE
4. MUSCULOSKELET.	<u>NAO</u> A NE	9. UROGENITAL <u>NAO</u> A NE
5. CIRCULATORY <u>NAO</u> A NE		10. OTHER <u>NAO</u> <u>(A)</u> NE

FEMORAL VESSELS: Right patent Left patent  
 WEIGHT (kg) 2.5 DATE 4/8/98 CURRENT TB TEST 4/1/98

**ABNORMAL FINDINGS:**  
(2) mild tartar & gingivitis.  
(10) normal uterus - deviates mildly to (L)

REPRODUCTIVE EVALUATION
UTERUS: NAO A NE ADHESIONS: MINOR MODERATE SEVERE PREGNANCY STATUS: PREGNANT: GL (mm)= _____ BPD (mm)= _____ FL (mm)= _____ E/FHR (bpm)= _____ Gest. Age (days) _____ NONPREGNANT: UTERINE SIZE _____ CONTOUR/SHAPE _____ GENDER: M F

REPRODUCTIVELY SOUND AREPRODUCTIVE RE-EVALUATE NOT EVALUATED  
 COMMENTS: EAR TAG # 27719 (L)  
TPR-WNL

OVERALL CONDITION: EXCELLENT GOOD FAIR POOR

RECOMMENDATION: I CERTIFY TO THE BEST OF MY KNOWLEDGE THAT THIS ANIMAL HAS BEEN EXAMINED AND IS:  
 SATISFACTORY FOR SHIPMENT COMMENT:  
 SATISFACTORY FOR PROJECT COMMENT:  
 OTHER COMMENT: initial and review  
 DATE 4/8/98 EXAMINING VETERINARIAN: \_\_\_\_\_

# CALIFORNIA REGIONAL PRIMATE RESEARCH CENTER

## ANIMAL ACQUISITION RECORD

### A. Filled out by Research Services:

MCY 30749  
Species and ID#

03/26/98  
Acquisition Date (M-D-Y)

Qu Location QU 4-4

Perdiem Payor CRX01/8794

Project Code CRX01/GLP10

Colony O

CRPRC Generation 00

Mother's ID#  
(if known) N/A

Father's ID#  
(if known) N/A

ISIS Birthplace:

Institution code  
(if domestic born) N/A

Geographic code  
(if wild-caught) N/A

ISIS Acquisition Source:  
Institution code 4511 13 9X1

Census Flags \_\_\_\_\_

Social Code IH

REMARKS:

RECORDED BY: \_\_\_\_\_

### B. From Quarantine Screen-In Physical:

Sex: M \_\_\_\_\_ F X

Previous Identification 27719

Date of Birth (if known) 6/10/91 (M-D-Y)

OR

Estimated Age \_\_\_\_\_ years \_\_\_\_\_ months

Comments:

VETERINARIAN: \_\_\_\_\_

DATE: \_\_\_\_\_

35749

California Primate Research Center

1

Animal Number								Page		
Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P.)	HYDRATION (G.F.P.)	STOOL (N,SS,L,B)	Observation	Init
3-26-98									RECEIVED INTO QU 4-4	MR
4-1-98	2.5	M	0	0	0				.3cc ket; Tattoo	JES
4/9/98	2.5								0.3ml ketamine given IM Qu screen - in PE: @ ear tag - 27719 mild tartar & gingivitis observed & uterus deviated mildly to the (L) A: satisfactory screen - in	(IB) MR
4-14-98	2.40	M	0	0	0				.3cc ket	MR
4-28-98	2.50	M	0	0	0				0.35 cc KET; 3ml serum, 2ml red top, 1ml CBC, stool, rectal samples	MR
5-13-98	2.40	M	0	0	0				.35 cc KET; Bled 10mls	MR
5-27-98	2.50	M	0	0	0				.4cc KET Malaria Tx: Mefloquine 25mg/kg via NGT	MR
6-10-98	2.5	M	0						.3cc ket	JES
7-2-98									.3cc ket released from QU #3846	JES
7/2/98	2.49								So: Quarantine screen and physical examination (B)	(B)
[REDACTED]										
[REDACTED]										
Malaria Tx: Mefloquine 25mg/kg via NGT										
7/2/98									GLP 10: Bled 2ml for CBC and Chem, 2ml urine collected by cystocentesis. 0.3ml ketamine IM. Moved to BB4004-39	PA

VACCINATED TO TANKS

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

730620.01

04681 (2/90)

Animal Number								Page
Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P.)	HYDRATION (G.F.P.)	STOOL (N,SS,L,B)*
							Observation	Init

7/3/98 GLP10: 0.4ml Ketamine IM for baseline radiographs PA

Fe dextran DOSE 0.13 IM Q 7d  
 DRUG AMT. ROUTE FREQ.  
 7/9 7/31  
 START END DAY  
 30749 4004 89  
 AN# LOC  
 ADD COMMENTS:

7-16-98	2.30								BL
7/30/98	2.38								8mg
7/15/98									GLP10: 0.17ml Ketamine IM. Bled 6ml. 0.17ml medetomidine and 0.25ml atropine IM for bone scan. 0.10ml Ketamine IM. 0.17ml atipamezole IM. Returned to home cage. PA
7/31/98									GLP10: 0.17 Ketamine and 0.17ml medetomidine IM for bone scan. 0.24ml atropine IM. 0.17ml atipamezole IM. Returned to home cage. PA
8/5/98									20 mg/kg IV Total Volume: 1.2 ml PA
8/12/98	2.37								GLP10: 0.17ml Ketamine IM. Bled 8ml. 0.17ml medetomidine and 0.24ml atropine IM for bone scan. 0.10ml Ketamine IM. 0.17ml atipamezole IM. Returned to home cage. PA
8/19/98								G G N	SO: BAR Re: poor app. No biscuits were left in cage. Scant stool was present. P Monitor PA
8/19/98									20 mg/kg IV Total Volume: 1.2 ml PA
8/21/98									GLP10: 0.17ml ketamine IM for bone scan. Returned to home cage. PA
8/25/98									To HO1333 for surgery PA
9/20/98									SO: BAR, gave 0.24 cc ketamine & PA

G = good, F = fair, P = poor  
 N = normal, SS = semi-solid, L = liquid, B = Bloody

730620.01

Animal Number								Page		
Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P.)	HYDRATION (G.F.P.)	STOOL (N,SS,L,B)	Observation	Init
8/26/98									0.24 cc atropine im, intubated,	

Oxymorphone 0.24 14 TID  
 DRUG DOSE AMT. ROUTE FREQ.  
 8/26 8/26 2  
 START END DAY  
 30749 1833:3  
 ANT LOC

ADD COMMENTS:

									placed cephalic catheter & collected 0.5 cc blood for CBC, performed bilateral ovariectomy & @ iliac west by (see surgery report)	
8/27/98						F G N			SO: BAR, sutures intact P. Discharge to normal	SB
8/27/98	2.39								GLP10: Received [redacted] 125 mg/kg via NGT from 8/27/98 to 9/9/98 Dose Volume: 12.0 ml. Bled 9.6 ml	PA
9/4/98						F G N			SO: BAR. On report for bloody stool - unconfirmed, stool appears normal. A: unconfirmed bloody stool - 1st report. P: Monitor.	SS
9/2/98									GLP10: Bled 2.5ml for post-5x estradiol - wk 1	PA
9/9/98	2.39								GLP10: Bled 2.5ml for post-5x estradiol - wk 2	PA
9/10/98									GLP10: Received [redacted] 125 mg/kg via NGT from 9/10/98 to 9/23/98 Dose Volume: 12.0 ml	PA
9/16/98									GLP10: Bled 2.5ml for post-5x estradiol - wk 3	PA
9/23/98									GLP10: Bled 2.5ml for post-5x estradiol - wk 4	PA
9/23/98	2.45								GLP10: Bled 8.4ml for pharmacokinetics - DAY 28	PA

730620.01

G = good, F = fair, P = poor  
 N = normal, SS = semi-solid, L = liquid, B = Bloody



MCY 30749

California Primate Research Center

4

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (N,SS,L,B)\*

Observation

Init

Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P)*	HYDRATION (G.F.P)*	STOOL (N,SS,L,B)*	Observation	Init
9/24/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 9/24/98 to 10/7/98 Dose Volume: 12.3 ml	PA
10/7/98	2.44									
10/8/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 10/8/98 to 10/21/98 Dose Volume: 12.2 ml	PA
10/22/98	2.41								GLP10: Received [REDACTED] 125 mg/kg via NGT from 10/22/98 to 11/4/98 Dose Volume: 12.1 ml	PA
11/4/98	2.49									PA
11/5/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 11/5/98 to 11/18/98 Dose Volume: 12.5 ml	PA
11/18/98	2.49									PA
11/19/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 11/19/98 to 12/2/98 Dose Volume: 12.5 ml	PA
12/2/98	2.49									PA
12/3/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 12/3/98 to 12/14/98 Dose Volume: 12.5 ml	PA
12/14/98	2.51									PA
12/17/98									GLP10: Received [REDACTED] 125 mg/kg via NGT from 12/17/98 to 12/30/98 Dose Volume: 12.6 ml	PA

730620.01

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

MCY 30749

California Primate Research Center

5

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (N,SS,L,B)\*

Observation

Init

12/30/98

2.52

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 12/31/98 to 1/13/99  
Dose Volume: 12.4 ml

AN

12/31/98

1/13/99

2.49

m  
⊙ P O P

0.24 cc ket Im TB tested

AN

1/14/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 1/14/99 to 1/27/99  
Dose Volume: 12.5 ml

AN

1/27/99

2.54

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 1/28/99 to 2/10/99  
Dose Volume: 12.7 ml

AN

1/28/99

2/10/99

2.49

PA

2/11/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 2/11/99 to 2/24/99  
Dose Volume: 12.5 ml

PA

2/18/99

GLP10: 0.12cc ket im, bld, anal @ 0.12cc  
Medetomidine and 0.25cc Atropine for bone  
Scan, 0.12cc Atipamezole, returned to home  
Cage

AN

2/24/99

2.51

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 2/25/99 to 3/10/99  
Dose Volume: 12.6 ml

AN

2/25/99

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

(P4681290)

730620.01

GLP10: 0.12cc ket im, bld, anal @ 0.12cc 2/21/99

California Primate Research Center

MS/ 30749

6

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (N,SS,L,B)\*

Observation

Init

Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P)*	HYDRATION (G.F.P)*	STOOL (N,SS,L,B)*	Observation	Init
3/10/99	2.55 2.87								GLP10: Received [redacted] 125 mg/kg via NGT from 3/10/99 to 3/24/99 Dose Volume: 12.6 ml	[Signature]
3/11/99										
3/24/99	2.53								GLP10: Received [redacted] 125 mg/kg via NGT from 3/25/99 to 4/7/99 Dose Volume: 12.7 ml	[Signature]
3/25/99										
4/7/99	2.52								GLP10: Received [redacted] 125 mg/kg via NGT from 4/8/99 to 4/21/99 Dose Volume: 12.6 ml	[Signature]
4/8/99										
4/21/99	2.93								GLP10: Received [redacted] 125 mg/kg via NGT from 4/22/99 to 5/5/99 Dose Volume: 12.7 ml	[Signature]
4/22/99										
5/5/99	2.51								GLP10: Received [redacted] 125 mg/kg via NGT from 5/6/99 to 5/19/99 Dose Volume: 12.6 ml	PA
5/6/99										
5/19/99	2.50								GLP10: Bled 8.4ml for 9 month (W1C39) pharmacokinetics.	PA
5/19/99										

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

730620.01

10 EF 5 hand [redacted] 2.55 [redacted] 3/10/99

MCJ 30749

California Primate Research Center

7

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (MSS.LB)\*

Observation

Init

5/20/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 5/20/99 to 6/2/99  
Dose Volume: 12.5 ml

6/2/99

2.48

PA

6/3/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 6/3/99 to 6/16/99  
Dose Volume: 12.4 ml

PA

6/16/99

2.49

o. sec ket in blood and S.B

PA

6/17/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 6/17/99 to 6/30/99  
Dose Volume: 12.5 ml

PA

6/30/99

2.48

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 7/1/99 to 7/14/99  
Dose Volume: 12.4 ml

PA

7/1/99

7/14/99

2.48

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 7/15/99 to 7/28/99  
Dose Volume: 12.4 ml

PA

7/15/99

7/28/99

2.49

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 7/29/99 to 8/10/99  
Dose Volume: 12.5 ml

PA

7/29/99

730620.01

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

MCY 30749

California Primate Research Center

8

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR. READING

48-HR. READING

72-HR. READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (N,SS,L,B)\*

Observation

Init

8/11/99

2.52

PA

8/12/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 8/12/99 to 8/25/99  
Dose Volume: 12.6 ml

PA

8/18/99

GLP10: 0.18ml Ketamine 1M, Bled 8.5ml, 0.18ml midazolam  
and 0.25ml atropine 1M for bone scan. 0.10ml Ketamine  
1M, 0.18ml atipamezole 1M, returned to home cage.

PA

8/19/99

P/G

So: BAR. Re: poor app. ear flare &  
A: prob. due to sedation yesterday.  
P: normal

RAW

8/25/99

2.46

PA

8/26/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 8/26/99 to 9/8/99  
Dose Volume: 12.3 ml

PA

8/31/99

Moved to H01002-3

PA

9-1-99

So: W/O: 37 34, 0.25ml ketamine / 0.25 ml  
Atropine im to induce, 0.25ml Oxy im post

RA

Oxy morphine 0.25 IM BID  
DRUG DOSE AMT. ROUTE FREQ.

9/1 9/2 2  
START END DAY  
30749 1402-3  
ANT LOC

ADD COMMENTS:

op  
Shave crest biopsy performed.  
Returned to 1402-3 for recovery.

SL

9/2/99

F/G/N

So: BAR; autumntact  
P: OK to home cage

PA

9/2/99

Moved to B64004-58

PA

9/8/99

2.43

730620.01

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = Bloody

MC 30749

California Primate Research Center

9

Animal Number

Page

Date

WEIGHT (KG)

1B TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P.)

HYDRATION (G.F.P.)

STOOL (N,SS,LB)\*

Observation

Init

9/9/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 9/1/99 to 9/22/99  
Dose Volume: 12.2 ml

[Signature]

9/22/99

2.44

RJ

9/23/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 9/23/99 to 10/6/99  
Dose Volume: 12.2 ml

RJ

10/6/99

2.44

PA

10/7/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 10/7/99 to 10/20/99  
Dose Volume: 12.2 ml

PA

10/20/99

2.39

PA

10/21/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 10/21/99 to 11/3/99  
Dose Volume: 12.0 ml

RJ

11/3/99

2.48

RJ

11/4/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 11/4/99 to 11/17/99  
Dose Volume: 12.4 ml

RJ

11/17/99

2.44

RJ

11/18/99

GLP10: Received [redacted]  
125 mg/kg via NGT  
from 11/18/99 to 12/1/99  
Dose Volume: 12.2 ml

RJ

11/24/99

GLP10: 14 ml IS col for 15 month (wk 4) plasma col kinetics 1/20

G = good, F = fair, P = poor

N = normal, SS = semi-solid, L = liquid, B = Bloody

730620.01

mcy 30749

California Primate Research Center

10

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G.F.P)\*

HYDRATION (G.F.P)\*

STOOL (N,SS,L,B)\*

Observation

Init

Date	WEIGHT (KG)	TB TEST	24-HR READING	48-HR READING	72-HR READING	APPETITE (G.F.P)*	HYDRATION (G.F.P)*	STOOL (N,SS,L,B)*	Observation	Init
12/1/99	2.45									RJ
12/2/99									GLP10: Received [redacted] 125 mg/kg via NGT from 12/2/99 to 12/15/99 Dose Volume: 12.3 ml	RJ
12/15/99	2.51									RJ
12/16/99									GLP10: Received [redacted] 125 mg/kg via NGT from 12/16/99 to 12/29/99 Dose Volume: 12.6 ml	RJ
12/29/99	2.54									RJ
12/30/99									GLP10: Received [redacted] 125 mg/kg via NGT from 12/30/99 to 1/12/00 Dose Volume: 12.7 ml	RJ
1/12/00	2.48									RJ
1/13/00									GLP10: Received [redacted] 125 mg/kg via NGT from 1/13/00 to 1/26/00 Dose Volume: 12.4 ml	RJ
1/24/00									GLP10: Bled 8.4ml for 17mark (WK 74) pharmacokinetics	PA
1/26/00	2.48									RJ
1/27/00									GLP10: Received [redacted] 125 mg/kg via NGT from 1/27/00 to 2/9/00 Dose Volume: 12.4 ml	RJ
2/9/00	2.44									RJ
2/10/00									GLP10: Received [redacted] 125 mg/kg via NGT from 2/10/00 to 2/23/00 Dose Volume: 12.2 ml	RJ

G = good, F = fair, P = poor  
N = normal, SS = semi-solid, L = liquid, B = bloody

730620.01

MCY 30749

California Primate Research Center

11

Animal Number

Page

Date

WEIGHT (KG)

TB TEST

24-HR READING

48-HR READING

72-HR READING

APPETITE (G,FP)\*

HYDRATION (G,FP)\*

STOOL (N,SS,L,B)\*

Observation

Init

2/14/00

GLP10: 0.17ml Ketamine/IM. Blood 6ml. 0.17ml medetomidine and 0.24ml atropine IM for bone scans. 0.17ml atipamezole IM. Returned to home cage.

PA

2/23/00

2.50

RJ

2/24/00

GLP10: Received [redacted] 125 mg/kg via NGT from 2/24/00 to 3/8/00 Dose Volume: 12.5 ml

PA

3/8/00

2.54

RJ

3/9/00

GLP10: Received [redacted] 125 mg/kg via NGT from 3/9/00 to 3/22/00 Dose Volume: 12.7 ml

RJ

3/22/00

2.53

RJ

730620.01

G = good, F = fair, P = poor, N = normal, SS = semi-solid, L = liquid, B = Bloody



05/01/98

17

TO: [REDACTED]  
PRIMATE CENTER

FROM: [REDACTED] EH&S Technician  
Animal Use and Care Administrative Advisory Committee

RE: Animal Care and Use Protocol #8048  
EFFECTS OF [REDACTED] ON OVARIECTOMIZED CYNOMOLGUS MONKEYS.

Your animal care and use protocol for the project shown above was reviewed by Animal Use and Care Administrative Advisory Committee on 04/30/98.

The protocol was approved by the committee as submitted.

This approval will remain in effect until: 04/30/99.  
Original approval date for this protocol: 04/30/98.  
Protocol may be continued by annual updates until: 04/29/01.

Federal laws and guidelines require that Institutional Animal Care and Use Committees review ongoing projects annually. For the first two years after initial approval of the protocol you will be asked to submit an annual update form, describing any changes in procedures or personnel. The committee may, at its discretion, extend approval of the project in one year increments until the third anniversary of the original approval of the project.

Approval may only be extended until the third anniversary of the original approval of the project. At that time, the protocol must be replaced by an entirely new submission.