programcontents :

Sponsors	2
Meetings and Events	8
Schedule	9
Wednesday	9
Thursday	9
Friday	11
Saturday	
Case Reports Abstracts	14
Other Presentation Abstracts	
Workshop Participants	
Record of Participation	40

2007 Board of Directors

President Buddy Capuano

Vice President Rick Lee

President-Elect Tom Nolan

Past President Jim Elliott Secretary Marion S. Ratterree Treasurer Stephen Curtis Trustees Pat Turner (2007) Ruth Woodward (2007) Iris Bolton (2008) Gwendalyn Maginnis (2008) Thanks to our sponsors for your continued support; without it, we could not have grown into the organization we are today.

APV Education Fund Elizabeth R. Griffin Research Foundation

Thank you to the Elizabeth R. Griffin Research Foundation for their continued grant support to APV's education fund. The Elizabeth R. Griffin Research Foundation, Inc. is a non-profit foundation that was established in 1999 in the memory of Elizabeth R. "Beth" Griffin (1975-1997). Beth, who was an honors graduate of Agnes Scott College, was working as a researcher at Yerkes Primate Center when she contracted the rare macaque-borne Herpes B virus and subsequently died. Following her death, Beth's family established the foundation with the fervent desire that the scientific community and general public could continue to learn from the life of a young woman who had so much to offer the world. The Foundation's motto is Safe Research Saves Lives. For more information about the Elizabeth R. Griffin Research Foundation, contact Jim Welch at (423) 612-7233.

Platinum Sponsors

BioReliance, Inc.

14920 Broschart Rd Rockville, MD 20850 Phone: (301) 610-2632 Fax: (301) 610-2590 www.bioreliance.com Contact: Kelton Chapman

BioReliance offers comprehensive animal diagnostic services for a wide variety of research animals, including rodents, rabbits, and simians. Our services include serological assessment, parasitology, bacteriology, virology, pathology, and histopathology. Our comprehensive animal health laboratory provides the range of consultative services necessary to monitor the health status of your animal colonies.

Bio-Serv

1 8th St, Ste 1 Frenchtown, NJ 08825 Phone: (908) 996-2155 Fax: (908) 996-4123 www.bio-serv.com Contact: Karen Froberg-Fejko

Bio-Serv has worked diligently to meet the ever-changing needs of the research community for more than 35 years. Our goal is to be your one source for experimental diets (standard and custom), medicated feed, medicated dosing tablets (MDs), liquid diets, Durtless Precsion Pellets[®], and enrichment treats, devices, and ingredients.

Charles River BRF

305 Almeda Genoa Rd. Houston, TX 77047 Phone: (713) 433-5846 Fax: (713) 433-6971 www.crl.com Contact: Tami Lass Andrus

BRF, a division of Charles River Laboratories, is the largest supplier of nonhuman primates. BRF is committed to providing an uninterrupted supply of captive bred and feral Cynomolgus monkeys of unmatched quality from Mauritius that are free of B virus, SRV, SIV, and other defects. BRF also provides customized technical services, quarantine facility, and boarding in group pens in state of-the-art facilities. Accredited by AAALAC, International since 1970, BRF is your ultimate resource for Mauritian Cynomolgus monkeys.

Covance Research Products

Jim Wells Country Rd #381 Alice, TX 78333 Phone: (361) 664-4984 Fax: (361) 664-8933 www.crpinc.com Contact: Karyn Armstrong Covance is a supplier of high-quality laboratory animals for biomedical research, including nonhuman primates, SPF rabbits, guinea pigs, and purpose-bred canines. Covance also provides custom antibody services.

Harlan

8520 Allison Pointe Blvd, Ste 400 Indianapolis, IN 46250 Phone: (800) 793-7287 Fax: (317) 806-6090 www.harlan.com

Harlan is a world leader in commercial production and supply of laboratory animals. Backed by more than 75 years of experience, Harlan offers 150 stocks and strains of laboratory animals, including mice, rats, hamsters, and rabbits. Harlan Teklad is a leader in the formulation of laboratory animal standard and custom research diets. Teklad ensures quality control by maintaining fixed formula manufacturing procedures and using quality ingredients. Teklad offers a complete diet line, as well as autoclavable, irradiated, and certified diets, plus bedding and cage liners.

Hoover's Transport

21319 Timber Ridge Dr Magnolia, TX 77355 Phone: (281) 259-0029 Fax: (281) 259-2299 Contact: Jim Hoover

Hoover's Transport provides quality animal transport services, and new this year, animal shipping crates. We offer extensive knowledge, based on 16 years of experience in importing, exporting, and domestic shipping. Our services have been used by animal providers, zoos, pharmaceutical companies, and research facilities.

Kritter Krates, Inc.

5533 Avanak Spring, TX 77389 Phone: (281) 288-0040 Fax: (281) 528-6161 Contact: Spencer R. Ellis

Kritter Krates specializes in the import procedures and the relocation of nonhuman primates. Kritter Krates also provides disposable shipping crates.

Merck & Co. Inc.

770 Sumneytown Pike, WP 44-201 West Point, PA 19486 Phone: (215) 652-5621 Fax: (215) 652-0999 www.merck.com Contact: Dale Frankenfield Merck & Co. is a global research-driven pharmaceutical company.

Pacific Forest Resources, Ltd

PO Box 67391 Chestnut Hill, MA 02467 Phone: (617) 879-0088 Fax: (617) 278-9142 www.pacificfr.com Contact: Jeffrey Rosenberg

Pacific Forest Resources provides Asian origin, captive-bred, tested pathogen-free, nonhuman primates to scientific research organizations world-wide. PFR's western trained veterinary staffs operate and manage the facilities. PFR offers customized holding and conditioning programs, as well as a full complement of supply, quarantine, holding, and transport services. PFR also sells blood products from its dedicated U.S. colony and new product registration services in China for companies interested in registering pharmaceutical, industrial, and agricultural substances with the Chinese regulatory authorities.

Primate Products, Inc.

34200 Doctors Hammock Rd. Immakalee, FL 34142 Phone: (239) 867-2020 Fax: (239) 867-2030 www.primateproducts.com

Contact: Paul Houghton

Primate Products, Inc. provides a full line of housing, handling, enrichment, and live animals to the research and education community. Our 9.0, 6.0, and 4.3 sq. ft. caging offers the latest in primate- and people-friendly design. Our handling system (pole and collar) has become the standard of the industry worldwide. We currently have in stock Indian and Chinese Rhesus; Cynos from Indonesia, Mauritius, Vietnam, and China; and African Green monkeys from St. Kitts.

workshop**sponsors**

Purina LabDiet

555 Maryville University Dr, Ste 500 St. Louis, MO 63141 Phone: (314) 317-5179 Fax: (314) 317-5281 www.labdiet.com www.testdiet.com Contact: George Nugent

Purina Mills is the international supplier of the LabDiet[®] and TestDiet[®] products used in lab animal nutrition. All products are manufactured in our ISO 9001:2000 certified plant in Richmond, IN. Diets are available in standard, certified, autoclavable, irradiated, vac-pak, and micro-pak. Custom diets are also available upon request.

Shared Enterprises, Ltd

P.O. Box 910 Richlandtown, PA 18955-0910 Phone: (215) 536-9388 Fax: (215) 529-7128 Contact: Michael Hsu

Shared Enterprises has been a leading supplier of Chinese cynomolgus and rhesus for many years. As a wholly owned, U.S.-based corporation, Shared Enterprises is dedicated to providing quality-defined Chinese macaques through a complete primate import quarantine service to the end-users. Using our own breeding and research facility, coupled with a network of reliable breeders/suppliers, Shared Enterprises is uniquely positioned to timely provide nonhuman primates and contract research/breeding services to the biopharmaceutical industry. Shared Enterprises remains committed to the ethical use of nonhuman primates.

VRL Laboratories

7540 Louis Pasteur, Ste 200 San Antonio, TX 78229 Phone: (877) 615-7275 Fax: (210) 615-7771 www.vrllabs.com Contact: Ron Berger

VRL offers simian diagnostic services and innovative solutions to customers worldwide. We have two testing locations, San Antonio, Texas and Suzhou, China.

Worldwide Primates, Inc.

PO Box 971279 Miami, FL 33197 Phone: (305) 378-9585 Fax: (305) 232-3838 Contact: Matt Block Worldwide Primates is an importer/distributor of nonhuman primates and operates a CDC-approved quarantine facility. Import/ export services and consultation is provided.

Gold Sponsor

R.C. Hartelust B.V.

PO Box 2170 Tilburg, N. Brabant 5049NL The Netherlands Phone: +31 13 4555189 Fax: +31 13 4550175 Contact: R.C. Hartelust This fully licensed company, established in 1960, is a major partner in the primate world. R.C. Hartlelust B.V. imports, quarantines, and exports all officially bred primates for the research world.

Suburban Surgical Company Inc.

275 Twelfth St Wheeling, IL 60090 Phone: (847) 537-9300 Fax: (847) 537-9061 www.subsurg.com Contact: Steve Kramer

For more than 60 years and three generations, we have designed and manufactured the highest-quality equipment available for the animal care market. Over the years, we have grown from a two-man operation to a family-owned and operated, full-service corporation with the staff and resources to meet the needs of this growing market. To continue to meet this commitment to quality, we have expanded again. Our new building has more than doubled our manufacturing, warehousing, and office space. The extra manufacturing capacity, warehousing capacity, and support staff allows us to continue to provide you with the quality equipment you need, when you want it, supported by the service you deserve. Whether you're outfitting a brand-new facility, remodeling, moving, or simply looking to add some new equipment to enhance your services, SSCI is dedicated to meeting your needs.

Silver Sponsor

Three Springs Scientific, Inc

1730 West Rock Rd Perkasie, PA 18944 Phone: (215) 257-6055 Fax: (215) 257-3773 Contact: Jim Sears

Three Springs Scientific, Inc. provides high-quality, purpose-bred, nonhuman primates; import quarantine; offsite housing in AAALAC-accredited facilities; temperature-controlled transportation; custom antibody production; and blood and serums for government, university, and private industry clients.

Bronze Sponsors

Allentown Caging

Route 526, PO Box 698 Allentown, NJ 08501-0698 Phone: (609) 259-7951 Fax: (609) 259-0449 www.allentowninc.com Contact: Cathy Gartland

Allentown Caging manufactures innovative animal cages and accessories, including standard and positive/negative control ventilated racks, air control products for integration with HVAC systems, fully sealed bio-containment ventilated housing, recirculating aquatic housing systems, biological work stations, thermoplastic micro-barrier cages, social interaction primate systems, plastic rabbit and guinea pig cages, kennels, and poultry housing.

Animal Specialties and Provisions, LLC

2400 Milford Square Pike Quakertown, PA 18951 Phone: (215) 804-0144 Fax: (215) 804-0145 www.animalspecialties.biz

workshop**sponsors**

Contact: William Clarke

Animal Specialties and Provisions is an ISO 9002-certified PMI LabDiet Dealer serving the Delaware Valley with laboratory animal diets and beddings, as well as facilities around the world with environmental enrichment resources. Our line for nonhuman primates addresses the need for variety, durability, safety, and economy. Working with facilities across the country has allowed us to develop enrichment items for all species incorporating valuable feedback from the animal care staff.

Beijing Prima Resources

83, Fuxing Rd, Bldg E11, Ste 132 Beijing, Beijing Phone: (267) 828-0120 Fax: +86-10-51607403 nhp.primaresources.com Contact: Weizhen Fang

Prima Resources is an industry leader in supplying Chinese, purpose-bred NHP and related services. Through the strategic alliances with most breeding farms in China and quarantine facilities in the U.S, Prima Resources has secured the unique position as a single-source supplier. Experienced professionals will oversee and facilitate the supply process from production to delivery, allowing us to provide the biomedical research community a continuous, reliable supply of Chinese premium purpose-bred NHP.

BSI-Brain & Software International, Inc

3495 Piedmont Rd NE, Bldg 11, Ste 710 Atlanta, GA 30305 Phone: (404) 495-5961 Fax: (404) 233-8625 www.bsi-usa.com Contact: Johnny Yan

BSI is a software development company providing an integrated animal and healtj records and protocol tracking software. PSM—Primates System Management is specifically designed for animal husbandry facilities and laboratories dealing with regulated large animal species and nonhuman primates.

workshop**sponsors**

Buckshire Corporation

2025 Ridge Rd Perkasie, PA 18944 Phone: (215) 257-0116 Fax: (215) 257-9329 Contact: Sharon Hursh

Buckshire is a licensed contract quarantine facility that provides import/export services to the research industry. There is a limited amount of short-term contract testing space available.

Chembio Diagnostic Systems

3661 Horseblock Rd Medford, NY 11763 Phone: (631) 924-1135 Fax: (631) 924-603 www.chembio.com Contact: Les Stutzman

Chembio develops and distributes rapid, easy-to-use, convenient, animal side serological assays for the detection of active TB in nonhuman primates (NHP) using a small aliquot (30 ul) of whole blood, serum, or plasma. The assay is USDA-licensed and is exclusively distributed throughout North America by Centaur. Precise results are obtained in 20 minutes or less and it is less traumatic to the animals. As a qualitative assay, no external equipment is required, enabling it to be used as an ideal field test for detection of TB in NHP.

Chinese Laboratory Primate Association

Rm A201, Bldg 16, Area 1, Anyuan Dongli Chaoyang District, Beijing, China Phone: +86-10-64896353 Fax: +86-10-64896353 www.cipa.org.cn

The Chinese Laboratory Primate Association (CLPA) consist of 30 NHP breeding farms and 20 entities specialized in research quarantine trading. It acts as a bridge between government and its members, assisting government in establishing a quota system and to supervise market. CLPA also develops academic exchanges among members and end-users, helps end-users to access high-quality animals, and provides assistance.

Frame's Animal Transportation

1119 Haverford Rd Ridley Park, PA 19078 Phone: (610) 399-5166 Fax: (610) 399-5165 Contact: Bill Frame

Frame's Animal Transportation provides care and transportation for live research animals in climate-controlled trucks nationwide. Our 40 years of experience assures the highest quality of service. Primate crates are custom built to USDA standards to meet your size requirements.

Instech Solomon

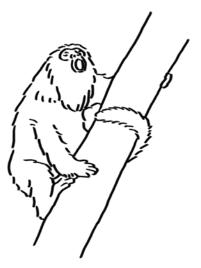
5209 Militia Hill Rd. Plymouth Meeting, PA 19462 Phone: (610) 941-0132 Fax: (610) 941-0134 www.instechlabs.com; www.solsci.com Contact: Tom Nolan

Instech Solomon was there at the beginning. In the late 1960s, Michael Loughnane developed a single-channel rat swivel. He started Instech Laboratories in 1971 to commercialize the swivel and other laboratory animal's infusion products. In 1983, Andrew Jacobson launched the Access Technologies VAP subcutaneous access ports. Jacobson's second customer was Tom Nolan, DVM at Merck Research Labs. Both started Solomon Scientific after Dr. Nolan retired from Merck. Paul Loughnane joined Instech in 1995. Now all four are together at Instech Solomon to deliver laboratory animal infusion systems for laboratory animals including nonhuman primates.

Lab Products, Inc.

11300 Rockville Pike Rockville, MD 20852 Phone: (888) 257-3532 Fax: (301) 468-3237 www.labproductsinc.com Contact: Betty Fatzie For more than 35 years, Lab Products has remained the world's leading innovator of laboratory animal housing and care equipment. We provide cost-effective solutions for housing animals, ranging from mice

to nonhuman primates, with the most efficient use of available area, time, and labor. Our product lines extend from classic conventional mouse cages to state-ofthe-art independently tested and certified AllerZone Micro-Isolators and Enviro-Gard Environmental Control Systems. We offer support with building HVAC design, room layouts, facility integration, 24-hour remote environmental monitoring/control, airflow balancing, allergen controls, equipment installation, training, and certification. Each system is designed to help ensure the well being of animals and personnel, to meet your research requirements, and to minimize experimental variables. Plus, Lab Products offers a unique alternative watering system, Hydropac[™] and biocontainment caging system.



The Mannheimer Foundation, Inc 20255 SW 360th St.

20255 SW 360th St. Homestead, FL 33034 Phone: (305) 245-1551 Fax: (305) 245-7650 Contact: Joseph L. Wagner

The Mannheimer Foundation, Inc. is a notfor-profit organization whose mission is the advancement of biomedical research directed towards the improvement of human and animal health. The Foundation maintains SPF breeding colonies of Cynomolgus macaques (SPF for SRV/D, SIV, STLV, and B-virus), Rhesus macaques, and Hamadryas baboons on our 82-acre main site in Miami. Support for research is provided at both of the Foundation's modern facilities, including advanced imaging, aseptic surgery, necropsy, and diagnostic laboratory capabilities. Biological tissues and body fluids are also available to support scientific research Nonhuman primate reproduction, genetics, behavior, and caging systems design are areas of ongoing Foundation research. Training opportunities are available for senior veterinary students and graduate veterinarians. The new 200-acre Haman Ranch facility is located 35 miles east of Ft. Myers, Florida.

Zoologix, Inc.

9811 Owensmouth Ave, Ste 4 Chatsworth, CA 91311 Phone: (818) 717-8880 Fax: (818) 717-8881 www.zoologix.com Contact: Steven Lloyd Zaologiy, parforma forthered

Zoologix performs fast, sensitive, specific PCR-based diagnostic testing for infectious diseases of primates and other animal species. See our assay menu, disease datasheets, specimen shipping instructions, and order form online.

: meetings&events

Board of Directors Meeting Wednesday, October 10, 7:00 p.m.–9:30 p.mBottec Saturday, October 13, 12:00–1:00 p.mBottec	elli Room elli Room	
Registration Wednesday, October 10, 3:00 p.m.–9:00 p.m. Thursday, October 11, 7:00 a.m.–5:00 p.m. Friday, October 12, 8:00 a.m.–12:00 p.m.	Foyer	
Speaker Ready Room Wednesday, October 10–Saturday, October 13DaVi	nci Room	
Primate Center Veterinarians Meeting (regional primate center vets only) Thursday, October 11, 5:30 p.m.–6:30 p.m		
Industry Veterinarians Meeting Thursday, October 11, 5:30 p.m.–6:30 p.m.	iice Room	
Hospitality Suite — hosted by the Hospitality Committee Wednesday, October 10, 7:00 p.m.–12:00 a.m	e 225–227	

WEDNESDAY	3:00 p.m9:00 p.m.	Registration (Foyer)
10.10.06	7:00 p.m.–9:30 p.m.	Board of Director's Meeting Bo ecelli Room
	7:00 p.m12:00 a.m.	Hospitality Suite Suite 225–227
THURSDAY 10.11.06	7:00 a.m.–8:00 a.m.	Continental Breakfast (Atrium
	7:00 a.m5:00 p.m.	Registration (Foyer)
	8:00 a.m.–8:15 a.m.	Welcome & Announcements: Buddy Capuano Venice Tuscany Ballroom
		Travel Award Presentations: Ruth Woodward <i>Venice Tuscany Ballroom</i>
	8:50 a.m9:40 a.m.	 Session 1 Case Reports: Novel Treatment Strategies Moderator: Pat Turner Venice Tuscany Ballroom Effects of the Macrolide Drug Tylosin on Chronic Diarrhea in Rhesus Macaques, Rebecca S. Blackwood[*] Treatment of Conjuctivitis by Subtenon Injection, Sylvia Gografe Using Fenbendazole in Commercial Non-Human Primate Diets and a Fecal Parasite Concentrator to Manage Intestinal Parasites in Macaques, Lorna Millen Asymptomatic Ependymal Cyst in Adult Cynomolgus Macaque, Kelly P. Yamada[*] Trichobezoar Prevention Using Soy Lecithin Supplementation in Galagos (Otolemur garne i), Margaret S. McTighe Urinary Alkalinization in the Treatment of Traumatic Rhabdomyolysis in Rhesus Macaques (Macaca mula a), Pablo R. Morales Localized Allergic Dermatitis Treated with Tacrolimus Ointment in a Rhesus Macaque (Macaca mula a), Stephanie Torreilles [*] Student Travel Award Recipient
	10:00 a.m.–10:10 a.m.	Break Prefunction Room

10:10 a.m.-12:10 p.m. Session 2 Case Reports: Emergency and Critical Care Moderator: Ruth Woodward Venice Tuscany Ballroom Acute Anorexia and Lethargy Associated with Pyrexia in an Adult Rhesus Macaque (Macaca mula a), Kelly M.S. Hugunir Fractures Associated with a Breeding Colony of SPF Cynomolgus Monkeys, Joe Wagner Stress Cardiomyopthy Following Severe Soft Tissue Trauma in a Chimpanzee (Pan troglodytes), Brigid V. Troan Calcinosis Cutis in Rhesus Macaques (Macaca *mula* a) Associated with Trauma, Joyce K. Cohen Adenocarcinoma in a Chimpanzee, Bill Satterfield Management of Iatrogenic Hypoglycemic Coma in a Rhesus Macaque (Macaca mula a), Christina Cruzen Psychosocial Stress and Diet Choice, Lynn A. Collura Acute Renal Failure Associated with Sepsis in Three Rhesus Macaques (Macaca mula a), Kirstin F. Barnhart A Student Travel Award Recipient 12:10 p.m.-1:20 p.m. Lunch Atrium 1:20 p.m.-3:20 p.m. Special Topic: Diagnostic 101: A Laboratory Users' Guide to Pathogen Testing in Nonhuman Primates: Joe Simmons, Nick Lerche, Bob Peters, Julia Hilliard Venice Tuscany Ballroom 3:20 p.m.-3:30 p.m. **Break** (*Prefunction* Room 3:30 p.m.-5:30 p.m. **Round-table Discussion: Clinical** Moderator: Gwen Maginnis

 Round-table Discussion: Management

 Moderator: Gale Galland

 Tuscany
 Ballroom

 • CDC NHP Import Update

 • TB Testing Update

 • USDA Update

 5:30 p.m.–6:30 p.m.

 Industry Veterinarians Meeting

 Venice
 Ballroom

*Venice Ballroom*Colitis in Macagues

Alopecia

Electronic Records and Database Software

	5:30 p.m.–6:30 p.m.	Primate Center Veterinarians Meeting (regional primate center vets only) <i>Tuscany Ballroom</i>
	5:30 p.m.–6:30 p.m.	Cocktails (cash bar)
	7:30 p.m.–9:00 p.m.	Dinner Florence and Milan Ballroom
	10:00 p.m.–12:00 a.m.	Hospitality Suite Suite
FRIDAY 10.12.06	6:30 a.m.–7:30 a.m.	Platinum Sponsors' Breakfast (board members and platinum sponsors only) Donatello Room
	7:00 a.m8:00 a.m.	Buffet Breakfast (Atrium
	8:00 a.m.–12:00 p.m.	Registration (Foyer)
	8:00 a.m.–8:05 a.m.	Announcements: Buddy Capuano Venice Tuscany Ballroom
	8:05 a.m.–9:50 a.m.	 Session 3 Case Reports: Cutting-Edge Techniques Moderator: Rusty Brady Venice Tuscany Ballroom Techniques for Treating Vaginal Prolapses, Joseph T. Newsome Determining Efficacy of Post-endoscopic Biopsy Treatment with Sucralfate on Appetite in Nonhuman Primates, Morgan L. Singletary Non-Invasive Blood Pressure Determination for Continuous Anaesthesia Monitoring in Nonhuman Primates, Barthel Schmelting Molecular Epidermicological Investigation of Simian Varicella Virus Infection in Two Immunocompromised Rhesus Macaques (Macaca mula a), Steven T. Shipley Some Advances from the Use of Macaques in Gene Therapy, Rebecca L. Grant Limitations of Measles Diagnostic Testing in Rhesus Macaques (Macaca mula a), Lisa C. Halliday Colonoscopy in Rhesus Macaques (Macaca mula a) – Principles, Procedures, and Problems, Bruce Bernacky
	9:50 a.m.–10:15 a.m.	Herpes B Update: Julia Hilliard Venice Tuscany Ballroom
	10:15 a.m.–10:30 a.m.	Break Break (Prefunction Room

10:30 a.m.–11:30 p.m.	 Session 4 Case Reports: What's Your Diagnosis? Moderator: Gwen Maginnis Venice Tuscany Ballroom Persistent Conjunctivitis in a Rhesus Macaque (Macaca mula a), Nancy L. Merrill Focal Pyogranulomatous Encephalitis in a Rhesus Macaque (Macaca mula a), Maria T. Zayas Failure to Thrive in a Cynomolgus Monkey (Macaca fascicularis), Melaney K. Gee Abdominal Mass in a Squirrel Monkey, Rusty Brady
11:30 p.m.–12:15 p.m.	 Round-table Discussion: Cardiomyopathy in Nonhuman Primates Venice Tuscany Ballroom Cardiac Disease in New World Monkeys Alfonso Gozalo Cardiomyopathy in Captive Chimpanzees (Pan troglodytes) D. Rick Lee and Michael Lammey
12:15 p.m.–1:30 p.m.	Lunch Atrium
1:30 p.m.–3:45 p.m.	 Session 5 Case Reports: General Topics Moderator: Tom Nolan Venice Tuscany Ballroom Irregular Cutaneous Scalp Mass in a Rhesus Macaque (Macaca mula a), Tara G. Ooms Physiological and Behavioral Effects of Social Introduction on Adult Male Rhesus Macaques (Macaca mula a), Lara A. Doyle Multiple Congenital Deformities Including Sternoschisis and Scoliosis in a Newborn Rhesus Macaque (Macaca mula a), Lauren Drew Martin Gestational Trophoblastic Disease in an Adult Female Cynomolgus Macaque (Macaca fascicularis), Carol Emerson Compliant Housing of Nonhuman Primates in Preclinical Safety Testing, Susanne Rensing Diagnosis and Treatment of Nephrotic Syndrome in a Male Captive Chimpanzee (Pan troglodytes), Michael Lammey Fluid-Filled Cyst in the Caudal Abdomen of a Rhesus Macaque (Macaca mula a), Jennifer Bacon Visceral and Neural Larva Migrans in Rhesus Macques (Macaca mula a), Alfonso Gozalo Husbandry and Veterinary Procedures for a BSL-3 Macaque Facility, Lisa C. Halliday
3:45 p.m4:00 p.m.	Break (Prefunction Room
4:00 p.m.–5:00 p.m.	APV Business Meeting (members only) <i>Venice Tuscany Ballroom</i>

	5:30 p.m.–6:30 a.m.	Cocktails (cash bar)
	6:30 p.m.–10:00 p.m.	Dinner and Auction Florence and Milan Ballroom
	10:00 p.m.–12:00 a.m.	Hospitality Suite Suite
SATURDAY 10.13.06	7:00 a.m.–8:00 a.m.	Continental Breakfast (Atrium
	8:00 a.m.–8:05 a.m.	Announcements: Buddy Capuano Venice Tuscany Ballroom
	8:05 a.m.–9:05 a.m.	Special Topic—The Dynamics of Endodontic Treatment: Mario Alves Venice Tuscany Ballroom
	9:05 a.m.–10:05 a.m.	 Session 6 Case Reports: Unusual Cases Moderator: Gwen Maginnis Venice Tuscany Ballroom New Innovations in Primary Housing for Great Apes, Paul Spurlock Invasive Primate Neuroscience: Is More Regulation or Better Management the Key to Success?, Paul Honess Surgical Placement of Vascular Access Ports in Common Marmosets, Lessions Learned, Mary Sauer Fundamentals of Vascular Access Ports in Nonhuman Primates, Jan Bernal
	10:05 a.m.–10:20 a.m.	Break (Prefunction Room
	10:20 a.m.–10:45 a.m.	Special Topic—NHP Breeding in China: the Status and Prospective: WZ Fang Venice Tuscany Ballroom
	10:45 a.m.–11:00 a.m.	AMP Update: Jacquie Calnan <i>Venice Tuscany Ballroom</i>
	11:00 a.m.–11:15 a.m.	Journal of Medical Primatology: Proceedings and Publications: Preston Marx Venice Tuscany Ballroom
	11:15 a.m.–11:45 a.m.	ILAR's International Primate Plan: Buddy Capuano <i>Venice Tuscany Ballroom</i>
	11:45 a.m.–12:00 p.m.	Closing Remarks Venice Tuscany Ballroom
	12:00 p.m.–2:00 p.m.	Board of Director's Meeting <i>Bo ecelli Room</i>
APV 35th Annual W	/orkshop • October 11–13	

APV 35th Annual Workshop • October 11-13, 2007 • Charlotte, NC

Case report abstracts listed chronologically by category

Novel Treatment Strategies

Effects of the Macrolide Drug Tylosin on Chronic Diarrhea in Rhesus Macaques (*Macaca mula a*)

RS Blackwood*, RP Tarara, KL Christe, A Spinner, NW Lerche

California National Primate Research Center, University of California, Davis, CA

Diarrhea is the most frequently encountered gastrointestinal disease in captive rhesus macaques. The precise pathogenic mechanisms underlying chronic diarrhea in nonhuman primates are not well understood, but a persistent inflammatory component has been strongly implicated. This study evaluated the inflammatory changes in the colon of affected macaques and assessed efficacy of a 10-day course of tylosin in a cohort of 21 animals with chronic diarrhea. Stool quality was evaluated daily and fecal consistency was scored. Colonoscopies were performed; biopsy samples were characterized histologically and assayed for expression of tumor necrosis factor alpha (TNF α) mRNA. Blood samples collected pre-, mid-, and post-treatment were assayed for C-reactive protein (CRP). Results indicate that 63% of animals receiving tylosin showed improvement in stool quality compared to 10% in the sham-treated group. Histologically, 82% of animals in the tylosin-treated group had a reduction in the severity of colonic lesions post-treatment, as opposed to 40% of animals in the sham group. There was no significant difference in TNF α mRNA in either tylosin- or sham-treated animals before or after treatment. CRP levels serially decreased in tylosin treated monkeys; the average post-treatment CRP value for tylosin-treated animals was $11.96 \pm 3.86 \,\mu$ g/ml compared to $26.48 \pm 4.86 \,\mu$ g/ml for sham-treated controls. In conclusion, tylosin significantly improved the fecal consistency score (FCS), significantly decreased colonic inflammation, and significantly decreased serum CRP levels post-treatment in rhesus macaques with chronic diarrhea.

Treatment of Conjunctivitis by Subtenon Injection

S Gografe*, KD Hansen

Vanderbilt University, OAWA, Nashville, TN

Conjunctivitis is a common occurrence in nonhuman primates. It can become a chronic disease, especially in older animals or animals with predisposing co-morbidities such as Diabetes mellitus type 2. Treatment is usually accomplished with instillation of an antibiotic ointment with or without a steroid at least twice a day. However, if the monkey is not trained to present for treatment, the ocular instillation of ophthalmic ointment can be very challenging. Very often the animal needs to be chemically restrained. Being anesthetized twice daily for several days can be difficult and exhausting for the animal. Furthermore, technicians need to be trained and reliable to administer the instillation correctly. Hence, the result of the treatment very often is not satisfactory. In the author's experience, relapse of the condition even with a perfect compliance in the treatment regimen is very common in diabetic animals when treated with ophthalmic ointment alone or a combination of ophthalmic ointment and systemic antibiotics. Therefore, a treatment technique adapted from human medicine has been employed with several rhesus macaques (*Macaca mula a*) that exhibited Diabetes mellitus type 2 or metabolic syndrome: 3 subtenon injections of an antibiotic drug combined with a local anesthetic, 48 h apart. The technique has not only proven very suc-

cessful in curing the conjunctivitis but is also easily learned by the technician and less stressful for the animal.

Using a Fecal Parasite Concentrator for the Detection of Intestinal Parasites in Rhesus (*Macaca mula a*) and Cynomolgus Macaques (*Macaca fascicularis*)

L Millen*, K Armstrong, M Cottingham, G Fleurie

Covance Research Products, Alice, TX

Intestinal parasites in macaques and many other animals are routinely diagnosed by the use of fecal flotation. Concentration techniques can optimize the detection of parasites by increasing the number of eggs recovered and by floating heavier eggs such as those of liver flukes (Fasciola sp.). When our facility decided to implement fecal concentration of all samples instead of using simple fecal flotation, we had to choose a specific concentration technique. Traditional concentration techniques involve filtering the feces through gauze into a centrifuge tube, centrifuging, then decanting and resuspending the sample before examination. However, we routinely process 200 fecal samples at a time at our facility, so this process was much too labor intensive to consider. We are now using the Midi Parasep Faecal Parasite Concentrator (DiaSys Europe) to diagnose intestinal parasites in our population. The advantages of this device over other concentration techniques are numerous and include ease of use, cleanliness, and reduction of reagents. The unit comes pre-loaded with buffered formalin and requires only the addition of feces and 2.0 ml ethyl acetate. Prepared concentrator units can be held at room temperature for 24 h before centrifugation. The risk of contaminating the centrifuge is eliminated because the concentrator unit is completely enclosed. We ran 111 samples in parallel using both traditional fecal flotation with magnesium sulfate (1.2 to 1.3 sg) and the Midi Parasep system. The Midi Parasep system recovered approximately 15% more parasites than MgSO₄ flotation and was far superior for the detection of *Fasciola*.

Use of Fenbendazole in Commercial Nonhuman Primate Diets for the Elimination of Intestinal Parasites

L Millen*, K Armstrong, M Cottingham, G Fleurie

Covance Research Products, Alice, TX

Two groups of cynomolgus macaques (Macaca fascicularis) entering quarantine were fed commercial diets containing fenbendazole to determine the efficacy of this method of anthelminthic administration. Animals were singly housed indoors, 60 animals/room, and were subject to standard CDC quarantine procedures. Pooled fecal samples were obtained on day 1 of quarantine. In the Chinese-origin animal samples, we found hookworm eggs, Trichuris sp. eggs, Balantidium colEasciola sp. eggs, and Ascaris sp. eggs; the Vietnamese-origin animals tested positive for Trichuris sp. only. Animals were fed biscuit-type feed containing 600 ppm fenbendazole twice a day for 5 d, at an amount sufficient for their caloric needs based on body weight. On day 6, the animals were placed on standard non-medicated feed. Fecal samples were rechecked on day 13, and all animals were negative for intestinal parasites. A separate animal group in outdoor corn crib-style housing with gravel bases were also fed the medicated feed for parasite elimination. Pooled fecal samples from two of the cribs were submitted for analysis. Both cribs tested positive for Ascaris sp.; Crib 2 also tested positive for Trichuris sp. These animals were fed medicated feed for 2 d; on day 3 they were moved to cribs in which the gravel base had been dug out and replaced with clean gravel. The animals were fed medicated feed for 5 d. On day 13, fecal samples were negative for intestinal parasites.

Asymptomatic Ependymal Cyst in an Adult Cynomolgus Macaque (Macaca fascicularis)

KP Yamada^{1,*}, SJ Popilskis², AP Garaté³, MJ Oglesbee⁴, KMD LaPerle³

¹Tri-Institutional Training Program in Laboratory Animal Medicine and Science: Memorial Sloan-Kettering Cancer Center, Rockefeller University and Weill Medical College of Cornell University, New York, NY; ²Research Animal Resource Center and ³Laboratory of Comparative Pathology, Memorial Sloan-Kettering Cancer Center, New York, NY; ⁴The Ohio State University, Department of Veterinary Biosciences, Columbus, OH

An asymptomatic naïve adult male cynomolgus macaque (*Macaca fascicularis*) underwent a routine pre-study MRI of the brain. The MRI revealed a diffuse, bilateral cystic lesion, involving the occipital cortex, whose content was isointense with cerebrospinal fluid (CSF). On necropsy, the cystic structures communicated with the caudal aspect of the lateral ventricles and contained clear fluid compatible with CSF by fluid analysis. Histopathology showed ciliated ependymal cells lining the cyst with focal areas of hemosiderophages within the cyst wall. This lesion is compatible with an ependymal cyst, previously reported only in humans and generally considered an asymptomatic developmental anomaly.

Trichobezoar Prevention Using Soy Lecithin Supplementation in Galagos (*Otolemur garne i*)

MS McTighe*, M Feurtado

Vanderbilt University, Nashville, TN

Trichobezoars (hairballs) have been identified as a source of weight loss and death in prosimians. Otolemur garne ii (galagos or bush babies) are avid groomers prone to hairball formation. Vanderbilt University maintains a colony of approximately 90 galagos (average daily census) for use in comparative research. Hairballs were detected in 16 animals over 6 y (1% to 4% of the colony). Treatment of hairballs consisted of surgical removal (gastrotomy) or use on a terminal protocol; 11 of the galagos underwent gastrotomy, and 5 were used in nonsurvival surgical protocols. The largest hairball removed was 40 g, removed from a female that developed 2 hairballs, 2 y apart. The average age of galagos at time of hairball detection was 1.8 y. Of the 16 affected animals, 11 (69%) were females and 5 (31%) were males. From 1999 to 2004, preventative measures included weekly treatment with oral laxative (Laxatone) plus 100% abdominal palpation at each cage change. In early 2005, we changed the diet from Purina Cat Chow to Purina Lab Diet 5280 (Ferret Diet), a nutritionally comparable diet. Extrapolating from a 2003 feline nutritional finding, soy lecithin (135 mg/kg), a fat emulsifier that prevents ingested hair from sticking together and thus eliminates hairball formation, was added to a weekly food supplement of cooked mashed sweet potatoes. Since this supplementation was initiated in January 2004, we have seen a dramatic reduction in hairballs, with no cases occurring from 2006 to present.

Urinary Alkalinization in the Treatment of Traumatic Rhabdomyolysis in Rhesus Macaques (*Macaca mula a*)

CA Matchett¹, A Saucedo¹, PR Morales^{1,*}, JL Wagner¹, A Garcia²

¹The Mannheimer Foundation, Homestead, FL; ²Yerkes National Primate Research Center, Atlanta, GA

A 15-y-old, 9.1-kg female rhesus macaque (*Macaca mula a*), housed in an outdoor breeding group, presented with severe crush wounds inflicted by cagemates. Physical exam revealed

a capillary refill time of 6 s, pale mucous membranes, and a heart rate of 192 BPM. Severe ecchymosis, edema, and lacerations were noted on the extremities, thorax, perineum, and face. Due to marked hypovolemia, an intravenous catheter could not be inserted. An intraosseous catheter was placed, and blood samples were collected. After the animal was secured to a restraint board, a urinary catheter was also inserted. Lab result values included BUN 52 mg/ dl, creatinine 3.7 mg/dl, phosphorus 12.7 mg/dl, CPK > 30,000 U/L, and WBC 18,000 /µl (mature neutrophilia and lymphopenia). A urine test strip indicated a pH of 6.0 and blood 4+, in the absence of hematuria. These strips cross-react with myoglobin, a hemoprotein found in muscle and associated with acute renal failure 48 to 72 h post-crush injury. Current medical literature suggests urinary alkalinization (to pH > 6.5) increases myoglobin byproduct solubility in urine, allowing renal excretion with minimal tubular damage. This principle has been demonstrated to increase survival in earthquake victims, who incur analogous injuries. Mindful of this observation, we initiated fluid therapy with 0.9% saline + 50 mEg/L of sodium bicarbonate; 20 ml/kg was infused over 30 min, followed by 30 ml/kg/h for a total volume of 110 ml/kg. Urine output was monitored to maintain 1 to 2 ml/kg/h and a pH above 6.5. For two subsequent days, intravenous fluids were administered at 20 ml/kg/h for 100 ml/kg/d. Bicarbonate was discontinued once myoglobinuria abated. The animal responded well and lab values returned within normal limits. Three additional macaques with traumatic rhabdomyolysis (CPK > 65,000 U/L in one case) have been successfully treated with this protocol, with no mortality. Urinary alkalinization, correction of metabolic acidosis, and the treatment of hyperkalemia associated with rhadomyolysis make bicarbonate a rational adjunct therapy for the condition in the rhesus macaque.

Localized Allergic Dermatitis Treated with Tacrolimus Ointment in a Rhesus Macaque (Macaca mula a)

S Torreilles*, D McClure, C Davis

Stanford University, Stanford, CA

An adult male rhesus macaque presented with pruritus and an exudative, inflammed, erythematous papule of approximately 1.5 cm in diameter under the chin. It was first treated daily with diluted Nolvasan and triple-antibiotic ointment resolving after 10 d. The lesion recurred several times over a 2-mo period. A punch biopsy revealed an allergic dermatitis. Treatment with Protopic (tacrolimus) ointment, an immunosuppressive drug, was initiated and appears successful at this time. Based on a literature review, this is the first report of immunosuppressive topical treatment of atopic dermatitis (commonly found in humans) in a rhesus macaque.

Emergency and Critical Care

Acute Anorexia and Lethargy Associated with Pyrexia in an Adult Rhesus Macaque (Macaca mula a)

KMS Hugunin*, LA Colby, KA Eaton, JE Wilkinson

Unit for Laboratory Animal Medicine, University of Michigan, Ann Arbor, MI

An 8-y-old male macaque presented with lethargy, anorexia, pyrexia, and swelling of a chronic IV catheter site. Diagnostics revealed azotemia, hyperphosphatemia, hyperkalemia, hypoalbuminemia, isosthenuria, hematuria, and proteinuria with elevated alkaline phosphatase. Despite treatment, he became edematous and increasingly depressed, and was euthanized.

Gross lesions included hepatomegaly with a thick, pale material associated with heart base vessels. Histopathology revealed fibrinosuppurative vasculitis and cellulitis with bacteria in the heart base and great vessels and bilateral membranoproliferative glomerulonephritis. Final cultures grew *Staphylococcus* aureus. It is hypothesized that the catheter infection induced a vasculitis that progressed to septicemia and a resultant glomerulonephritis.

Fractures Associated with a Breeding Colony of SPF Cynomolgus Monkeys

JL Wagner*, A Saucedo, K Rivas, S Howell, M Daniel, K Breen, M Serrano, J Larin

The Mannheimer Foundation, Homestead, FL

This report describes the incidence of fractures in young male and female cynomolgus macaques in our SPF breeding colony. Animals housed in newly formed breeding groups often exert extraordinary physical activity during breeding. Unique fractures were apparent among females soon after introduction to males. Clinical history, radiographs and dexascan information will be presented. Results suggest careful attention to females upon introduction to new potential breeding partners.

Stress Cardiomyopathy Following Severe Soft Tissue Trauma in a Chimpanzee (*Pan trog-lodytes*)

BV Troan*

North Carolina Zoological Park/North Carolina State University College of Veterinary Medicine, Asheboro, NC

A 7-y-old male chimpanzee was presented after sustaining multiple bite wounds over the body during introduction to a new troop. Aggressive treatment was initiated including wound care, nutritional support, fluids, analgesics, antibiotics, and antifungals. The clinical condition initially worsened, with the development of acute renal failure and several deeper wounds becoming infected. However, blood work parameters and the animal's attitude began to improve over the next 8 to 10 d. On day 13, the chimpanzee went into cardiac arrest during immobilization. He was successfully resuscitated, but 2 d later arrested again during an immobilization procedure and died despite CPR and defibrillation. The major necropsy finding was heart failure due to severe cardiomyopathy characterized by subacute myocardial necrosis with contraction bands, regeneration, and early fibrosis. Lesions were most prominent in the papillary muscle and on the subjacent epicardial and endocardial surfaces. Other evidence of heart failure included renal nephrosis, acute pulmonary edema, and centrilobular hepatocellular atrophy. Multiple skin wounds were also observed. The clinical history and necropsy findings are consistent with stress cardiomyopathy (alternatively called brain-heart syndrome) in which myocardial necrosis follows an episode of acute, severe sympathetic stimulation. Classically, this syndrome is described in cases of brain trauma, but similar cardiac lesions have been reported after severe soft tissue injury.

Calcinosis Cutis in Rhesus Macaques (Macaca mula a) Associated with Trauma

JK Cohen*, P Sharma-Reddy

Yerkes National Primate Research Center, Lawrenceville, GA

A 3-y-old male rhesus macaque presented with chronic intermittent swelling of both hands. Upon examination, a white, chalky substance was expressible. The animal was treated with antibiotics (Rocephin) and anti-inflammatories (Metacam) along with surgical debulking. A

bacterial culture was negative. Radiographs revealed mineralized material present throughout both hands. Hematology and serum chemistry were non-remarkable. Ionized calcium was within normal limits (1.35). Urinalysis revealed a mild hyposthenuria (USG 1.005). An ACTH stimulation was performed (cortisol pre 22.1, post 39.6) which did not rule out hyperadrenocorticism. The swelling progressed despite treatment, and the animal was euthanized due to the severity of the condition. Necropsy and histopathology findings were consistent with calcinosis cutis but did not support a diagnosis of hyperadrenocorticism. There are few case reports of calcinosis cutis in nonhuman primates. Nine cases of calcinosis cutis have been noted in rhesus macaques at the Yerkes National Primate Research Center over 17 years. These cases of calcinosis cutis have occurred in extremities (digits and tails) and appear to be associated with a history of trauma at the site of formation. There does not appear to be an association between calcinosis cutis and elevated serum levels of calcium or any other serum chemistry abnormalities. Calcinosis cutis can be either metastatic or dystrophic and has been associated with hypercalcemia from underlying conditions such as neoplasia, hyperadrenocorticism or iatrogenic. Calcinosis cutis caused by dystrophic calcification has been reported in humans associated with immunodeficiencies, congenital disorders, fat necrosis, intrauterine herpes infection in neonates, and trauma.

Adenocarcinoma in a Chimpanzee

B Satterfield*, S Buchl, M Miller, M McArthur, L Ginsburg, K Barnhart

Michale E. Keeling Center for Comparative Medicine and Research, Bastrop, TX

Chimpanzees are used as a model for specialized areas of biomedical research because of their unique immunologic and genetic similarity to humans. This model is usually reserved for research projects when rodents or other nonhuman primates will not be predictive outcomes for the biomedical applications in humans. Carcinomas are not a commonly reported or diagnosed malignancy in chimpanzees. This case report will present in detail the clinical history of this case, including diagnostic imaging, biopsy techniques, and surgery of a salivary adenocarcinoma.

Management of Iatrogenic Hypoglycemic Coma in a Rhesus Macaque (Macaca mula a)

C Cruzen*

Wisconsin National Primate Research Center, Madison, WI

Although accidental overdoses of insulin in diabetic pets are not uncommon, reports of insulin overdose in nondiabetic animals are infrequent, and recommendations for treatment are difficult to find. In all reported cases, treatment begins very early after insulin administration (prior to the onset of hypoglycemic coma). In addition, the pharmacokinetics of high doses of glargine insulin is not well established. In our facility, a case of iatrogenic glargine insulin overdose leading to a comatose state was successfully managed with continuous administration of intravenous dextrose solution and supportive care. A 6-y-old female macaque being treated for a skin wound presented early in the morning as recumbent and non-responsive in her cage. Physical exam findings at presentation showed severe bradycardia and arrhythmia, shallow and slow respirations, hypothermia, dorsoflexion of neck, and rigid extension of legs. I-stat analysis revealed profound hypoglycemia and hypokalemia. Investigation revealed that the animal was inadvertently given 140 units of glargine insulin the previous morning. Intravenous fluids with added dextrose combined with frequent dextrose boluses as needed (at 12.5%, 25%, or 50%) were provided for 32 h, at which point the animal was able to sit up for

brief periods and was attempting to eat. Blood glucose and other vital signs remained stable after the animal was able to eat, but the animal remained lethargic, mildly ataxic, and blind for 72-96 h after initial presentation. Complete recovery of the animal, including resolution of blindness, occurred after 96 h.

Psychosocial Stress and Diet Choice

LA Collura^{1,2,*}, J Fisher², ME Wilson²

¹Division of Animal Resources, School of Medicine, and ²Yerkes National Primate Research Center, Emory University, Atlanta, GA

Obesity is a growing medical and societal concern in developed countries. While availability of inexpensive, calorically dense foods contributes to this epidemic, emerging data, largely from rodents, suggests exposure to chronic stressors changes food preferences and the consumption of "comfort foods." Because stress-induced increases in endogenous opioids can increase feeding, the present study tested the hypothesis that an opioid receptor antagonist would decrease consumption of a high-fat diet (HFD) in animals exposed to psychosocial stressors. As social subordination in stable groups of rhesus monkeys (Macaca mulatta) is an ethologically relevant psychosocial stressor, we quantified the intake of a low-fat diet (LFD) and HFD 24 h each day for 21 d in 2 groups of adult females by use of an automated feeding station that identified individual monkeys from a unique microchip in the carpal regions. Animals were treated with a 21-d extended release formulation of the opioid antagonist naltrexone or received no implant as the control condition. The order of treatment was randomized and was separated by a 21-d washout. Subordinate females consumed significantly more of both the LFD and HFD throughout the day and night during both control and naltrexone treatments. Consumption of these diets was restricted to the daytime for dominant females. Food intake was not associated with differences in plasma ghrelin. Subordinate females showed higher gains in both body weight and BMI. These data suggest that changes on reward pathways other than opioids may mediate stress-induced consumption of comfort foods.

Acute Renal Failure Associated with Sepsis in Three Rhesus Macaques (Macaca mula a)

KF Barnhart*, BJ Bernacky

MD Anderson Cancer Center, Bastrop, TX

One male and two female rhesus macaques, aged 7 to 18 y, were diagnosed with acute renal failure (ARF) and inflammation due to an unknown etiology. The initial presentation of these monkeys varied from lameness and diarrhea to retention of a dead fetus. Initial clinicopathologic findings were similar in all cases and included severe azotemia, hyperphosphatemia, hypertriglyceridemia, hyponatremia, hypokalemia, hypochloremia, decreased antithrombin III, and neutrophilia with a regenerative left shift. Urine output appeared decreased in all three cases, and in one case gross hematuria was noted. Initial treatment included intravenous fluids, antibiotics, Lasix, and nutritional support. Despite aggressive therapy and parenteral administration of Baytril and Imipenem to the first affected monkey, the animal's clinical condition deteriorated and the neutrophilia persisted. Although hepatic involvement was not evident, the severity of the ARF associated with oliguria and marked hypokalemia were consistent with Leptospirosis and penicillin was administered. A marked decrease in the neutrophil count in the first 24 h was noted; however, the monkey worsened clinically, developed signs of disseminated intravascular coagulation (DIC), and was euthanized. A necropsy was performed. Histopathologic examination confirmed a cortical nephropathy affecting primar-

ily the tubules and DIC. Bacterial cultures of the blood and kidneys were negative. Warthin-Starry staining of a kidney section identified spirochetes in the renal tubules; however, PCR analysis and immunohistochemical staining for *Leptospira* spp. were negative. In the two subsequent cases, earlier administration of penicillin was associated with a more favorable response and both monkeys recovered following approximately 1 wk of hospitalization and intensive care.

Cutting-edge Techniques

Techniques for Treating Vaginal Prolapses

RA Wagner, JT Newsome*

University of Pittsburgh, Pittsburgh, PA

This presentation will review typical pathogenesis associated with vaginal prolapse in macaques as well as historic and novel treatment modalities. An initial review of predisposing factors for vaginal prolapse and possible preventive medicine management issues will be reviewed. Clinical management and discussion concerning choices for medical remediation and or surgical repair options. Non-surgical techniques, including both mechanical and pharmacological interventions, will be reviewed. In addition, a review and update of surgical options including Caslick, colpocleisis, and sarcolpopexy procedures in relation to case studies will be presented.

Determining Efficacy of Post-endoscopic Biopsy Treatment with Sucralfate on Appetite in Nonhuman Primates

ML Singletary*

Tulane National Primate Research Center, Covington, LA

Endoscopy with duodenal biopsy is a diagnostic tool frequently used for clinical and research purposes in nonhuman primates. Although many institutions use this procedure, post-procedural treatments are often varied. We performed a study using animals in which endoscopy with duodenal biopsies were completed as requested in the IACUC-approved protocol to which the animals were assigned. After endoscopy was performed, animals were randomly separated into 2 groups; the first group received sucralfate during the procedure and for 5 d post-procedure, and the second group received no sucralfate. This presentation will include the results of this study and an overview of the mechanism of action of sucralfate. A review of current literature in regards to the pharmacokinetics of the drug, its proven uses, and interactions with growth factors present at duodenal ulcer sites will also be discussed.

Non-invasive Blood Pressure Determination for Continuous Anesthesia Monitoring in Nonhuman Primates

B Schmelting^{1,*}, B Egner², SH Korte¹, G Weinbauer¹

 $^1\!\mathrm{Covance}$ Laboratories GmbH, Muenster, Germany; $^2\!\mathrm{S+B}$ medVet GmbH, Babenhausen, Germany

Pivotal and serious side effects of anaesthetic drugs are depression of respiration and of the cardiovascular system, while increased blood pressure (BP) of $\geq 10\%$ during anesthesia is considered to be a positive response to nociceptive stimulation that may require appropriate

action as well. Invasive techniques represent the major approach for accurate blood pressure determination in small species obviating routine use in veterinary practice. Non-invasive high-definition oscillometry allows visible, real-time screen control of measurements using BP amplitude scans up to 16.000 Hz, and was used for anesthesia monitoring in Callithrix jacchus (n = 6) and Macaca fasciculari(m = 6) under 2 different anesthetic regimens: ketamine (cynomolgus: 10 mg/kg, marmoset: 50 mg/kg) and ketamine-xylazine (cynomolgus: ketamine 10 mg/kg, xylazine 0.3 to 0.7 mg/kg; marmoset: ketamine 15 mg/kg, xylazine 3 mg/kg). Systolic and diastolic BP, mean arterial pressure (MAP) and pulse/min were determined individually in 1-min intervals. After ketamine injection, initial increases in pulse rate and MAP were only observed in marmosets, whereas both species showed slightly decreased BP and pulse rate afterwards. Following ketamine-xylazine injection, MAP reduction was more prominent and heart rate decreased up to factor 2 (cynomolgus) or 3 (marmoset). Despite a higher dose, xylazine appeared better tolerated by marmosets, as the lowest MAP (approximately 45 mm Hg) was noted in cynomolgus monkeys. The cardiovascular changes remained during recovery. Our findings demonstrate the feasibility and convenience of non-invasive blood pressure determination for continuous anaesthesia monitoring in nonhuman primates.

Molecular Epidemiological Investigation of Simian Varicella Virus Infection in Two Immunocompromised Rhesus Macaques (*Macaca mula a*)

ST Shipley*, K Kolappaswamy, R Mahalingam, D Gilden, BK Kleinschmidt-DeMasters, LJ DeTolla

University of Maryland , Baltimore, MD

We describe a clinical and epidemiological investigation of two cases of simian varicella virus (SVV) infection in immunosuppressed rhesus macaques housed in a single animal room. The two macaques developed acute generalized maculo-papular lesions on the skin, postgamma irradiation. At necropsy, following euthanasia for humane reasons, both animals had acute inflammatory lesions on the lungs and petechiations on the GI tract, and one had white raised plaques of approximately 1 mm diameter multifocally on all liver lobes. On the basis of real-time DNA PCR and histopathology, a confirmatory diagnosis of SVV was made. As a part of the epidemiological investigation, real-time DNA PCR was also performed on saliva samples from the rest of the animals in the room. SVV DNA was found in saliva from 2 animals which were not immunocompromised. Subclinical reactivation of SVV and shedding of virus through saliva could be considered the source of infection of these immunocompromised macaques.

Some Advances from the Use of Macaques in Gene Therapy

RL Grant*

University of Pennsylvania, Philadelphia, PA

Macaques have become the animal model of choice to analyze our viral gene transfer vectors before they are used in human clinical trials. Although our viral gene transfer vectors are extensively tested in murine models, murine species do not represent as close a biological and physiological model to the human as do macaques. Additionally, the toxicological and immune responses of a macaque more closely represent that of humans. The viruses we used to create replication-defective gene transfer vectors are retroviruses, adenoviruses or adeno-associated viruses. Retroviruses are a class of enveloped viruses that contain a single-stranded RNA molecule as the genome. Subsequent to infection, the viral genome is reverse transcribed into double-stranded DNA, which integrates into the host genome and is expressed as proteins. Adenovirus vectors can infect non-dividing cells with high efficiency. Unlike retroviral vectors, they do not integrate into the host genome; thus, gene expression is lost with time. Adeno-associated viruses are human parvoviruses that are dependent on a helper virus, usually an adenovirus, to proliferate.

Limitations of Measles Diagnostic Testing in Rhesus Macaques (Macaca mula a)

LC Halliday*, K Mansfield, JH Simmons, JD Fortman

University of Illinois at Chicago, Chicago, IL

Three juvenile male rhesus macaques from a single commercial source presented during the quarantine period with skin lesions consistent with measles infection, including maculopapular rash and respiratory disease. The animals were imported from China in late 2006 and completed primary quarantine without incident. One animal arrived at the University of Illinois at Chicago in a group of 10 juvenile and 8 adult rhesus in January 2007. The other 2 animals arrived in a group of 15 juvenile rhesus in February 2007. All 3 animals were euthanized due to significant respiratory compromise approximately 4 to 6 wk after arrival. Although clinical signs were consistent with measles, limitations in current diagnostic testing did not allow a definitive diagnosis to be made antemortem. Blood samples were submitted to commercial laboratories for serologic and polymerase chain reaction (PCR) testing for measles virus; results from ELISA, MFIA, and PCR were negative. The diagnosis was not confirmed until tissues collected at necropsy were submitted for immunohistochemistry, which showed they demonstrated high levels of measles antigen in multiple tissues. However, serum submitted after necropsy tested positive for measles virus antibody by immunofluorescence assay. This presentation will focus on the limitations of current commercial serologic and PCR testing encountered during diagnosis of an outbreak of measles in recently imported juvenile Chineseorigin rhesus macaques. In addition to the diagnostic testing procedures, an overview of the clinical course of disease in these animals will be provided.

Colonoscopy in Rhesus Macaques (Macaca mula a): Principles, Procedures, and Problems

B Bernacky*, K Barnhart, C Baron, R Rodriguez, M McArthur

MD Anderson Cancer Center, Bastrop, TX

For diagnosis of large intestinal disease, endoscopic evaluation of the colon using a flexible endoscope can be of tremendous scientific value and routinely performed in rhesus macaques. To accomplish this, certain criteria must be met to ensure success: a thorough knowledge of the equipment, technique and animal anatomy involved; proper patient preparation; the ability to obtain adequate biopsy samples; and the use of personnel in defined roles. This presentation will describe the indications for performing a colonoscopy, the required equipment, personnel needed, patient preparation, technique, normal findings, the complications inherent to performing a colonoscopy, and those specific to colonoscopy in macaques.

What's Your Diagnosis?

Persistent Conjunctivitis in a Rhesus Macaque (Macaca mula a)

NL Merrill*

APV 35th Annual Workshop • October 11-13, 2007 • Charlotte, NC

Madigan Army Medical Center, Steilacoom, WA

From August 2004 to February 2005, an 8-y-old male rhesus macaque had recurring problems of the left eye that were initially diagnosed as conjunctivitis; the eye was swollen shut, the eyelid was reddened, and a watery drainage was present. The macaque had been housed at Walter Reed Army Institute of Research for 2 y without any medical problems except for GI upset, was singly caged, and had always been Tb negative. Multiple evaluations, cultures, and treatments (antibiotics and steroids) were tried without complete resolution of the eye problems. After 6 mo, a foreign object was discovered in the eye; it had been placed and sutured (non-resorbable 5"0") behind the left eyeball during an ophthalmic protocol in 1998. According to the records, this object had been removed in December 2000. It took 4 y for the suture to degrade and allow the plastic tubing to migrate and cause problems.Had these conjunctivitis symptoms arisen during a routine Tb test, the animal would have been euthanized.

Focal Pyogranulomatous Encephalitis in a Rhesus Macaque (Macaca mula a)

M Zayas*, E Arifin, R Young, ML Voytko

Wake Forest University, Winston-Salem, NC

A 20-y-old female rhesus macaque developed a left-side head tilt with contralateral facial paralysis and proprioceptive deficits. Clinical history included an injury to the right ear resulting in a fistulous tract below the right pinna with subsequent obliteration of the associated ear canal. More recently, possible head trauma occurred during recovery from ketamine-induced anesthesia. Corticosteroids and a 7-d course of antibiotic therapy resulted in temporary cessation of the clinical signs. These signs reoccurred 1 wk later, with worsening of the previous signs. The same treatments were reinstated and again the clinical signs abated. Magnetic resonance imaging (MRI) indicated a mass lesion on the right side involving the brain stem and the adjacent cerebellum. Complete blood count (CBC) demonstrated a mild neutrophilia, and stool samples revealed occult fecal blood. Computed tomography (CT) did not disclose abnormalities of the abdominal organs. The animal was euthanized because of the deteriorated clinical condition and poor prognosis. The brain was preserved intact and, upon sectioning, a well-demarcated, irregularly shaped, multilobular, pale yellow mass was identified in the cranial part of the brain stem with compression of the adjacent cerebellum and displacement of fourth ventricle, in association with a 1-mm diameter bony mass arising from the right cranial vault. Histologically, the lesion was diagnosed as pyogranulomatous inflammation and the bony mass was diagnosed as focal chronic proliferative inflammation. Periodic acid Schiff and Gram stains failed to identify causative agents. Immunohistochemistry for amoebae were negative, and electron microscopy provided no further information as to etiology.

Failure to Thrive in a Cynomolgus Monkey (Macaca fascicularis)

MK Gee*, E Arifin, JM Cline, AJ Bennett

Wake Forest University Health Sciences, Winston-Salem, NC

A 3.5-y-old male cynomolgus monkey was examined for intermittent diarrhea, abdominal distension, and failure to thrive in comparison to age-matched cage mates. Routine diagnostics included complete blood count, serum chemistry, urinalysis, rectal cultures, fecal exams for parasite ova and giardia, survey radiographs, and abdominal ultrasound. Results of these tests revealed mild to moderate anemia and thrombocytosis, and moderate hypoproteinemia and hypoalbuminemia. Polymerase chain reaction assay for simian retrovirus (SRV) types 1 and 2 were negative. Serum hormone levels for testosterone, growth hormone, insulin-like

growth factor-1, and thyroxine (T4) were within normal range and were used to rule out hypothyroidism and pituitary dwarfism. Pre- and post-prandial bile acid values excluded congenital and acquired portosystemic shunt. Negative titers for anti-endomysial and anti-tissue transglutaminase antibodies, along with a gluten-free diet, were used to rule out gluten-sensitive enteropathy. Serum B12 and folate levels were lower than normal values referenced for humans, dogs, or cats, and were suggestive of exocrine pancreatic insufficiency, though fecal trypsin enzymatic activity was present and only a marginal response to dietary supplement with pancreatic enzymes was noted. No diagnosis was confirmed, and the condition did not improve. Humane euthanasia was elected. Gross morphologic and histologic necropsy findings revealed a persistent left superior vena cava, premature physeal closure, and a systemic chronic inflammatory condition. A genetic basis for the multi-systemic abnormality of growth was suspected.

Abdominal Mass in a Squirrel Monkey

SV Gibson¹, A Brady^{1*}, JC Ruiz², CR Abee²

¹Department of Comparative Medicine, University of South Alabama, Mobile, AL; ²Michale E Keeling Center for Comparative Medicine and Research. Bastrop, TX

In February, an aged, wild-caught, Guyanese female squirrel monkey, housed in a small group breeding run, was discovered with a distended abdomen. She was hypothermic and lethargic. An enlarged uterus consistent with pregnancy was palpated. Ultrasonography revealed an abdominal mass but no fetus. Absolute leukopenia (WBC = 2.18 m/mm³) with moderate normocytic, normochromic anemia (HCT = 22.9) was detected in the complete blood count. Serum chemistry abnormalities included hypoproteinemia (TP = 5.2 g/dl), hypocalcemia (CA = 7.3 mg/dl), mild azotemia (BUN = 43 mg/dl), and hyperphosphatemia (PHOS = 9.2 mg/ dl). Supportive therapy was initiated and exploratory surgery undertaken to determine the etiology of the mass. A large, multinodular mass arising from the uterus and adhering to the omentum was found. The surgeon determined that the mass was inoperable. Because of her age, condition, and poor prognosis, the monkey was euthanatized. At necropsy, the uterus was markedly enlarged, with multiple nodules extending from the myometrium. The uterine lumen contained dry white exudate, and the endometrium was effaced. Small raised white foci and nodules were found within the omentum, mesentery, and the dorsum of the liver. Two small cysts were found on the dorsum of the caudal lung lobes. Microscopically, there were multiple large areas of coagulative necrosis with multifocal bacterial colonies and locally extensive suppurative inflammation. The myometrium was highly cellular, with largely ovoid cells with marked nuclear pleomorphism and abundant clear to lightly eosinophilic cytoplasm.

General Topics

Irregular Cutaneous Scalp Mass in a Rhesus Macaque (Macaca mula a)

TG Ooms*, EP Ribka

Tulane National Primate Research Center, Covington, LA

A 21-y-old, 7.4-kg female rhesus macaque (*Macaca mula a*) was presented for apparent trauma to the head. This macaque was born at the Tulane National Primate Research Center and maintained on a globoid cell leukodystrophy breeding protocol; polymerase chain reaction confirmed carrier status for this autosomal recessive genetic disorder. Viral screening was

negative for all retroviruses but positive for Cercopithecine herpes virus-1. Examination revealed severe dental disease and a 3 cm x 3 cm x 1.5 cm irregularly shaped, lobulated, narrow based mass medial and caudal to the dorsal aspect of the right pinna. The surface of the mass was ulcerated. A complete blood count and serum chemistry were within normal limits. One week after presentation, the mass was excised with blunt and sharp dissection to elevate and remove the mass. The mass was confined to skin, which was closed after the excision; the animal recovered routinely. Histopathology revealed a circumscribed mass composed of ribbons, cords, and tubular formations of basal epithelial cells. The stroma was composed of dense fibrous bands with small mucinous zones and mildly irregularly distributed chronic inflammation, consistent with a basal cell tumor with a garland pattern. Basal cell tumors are very common skin tumors in the human population; most tumors are confined to the face, ears, neck, scalp, shoulders, or back, and have been linked to previous sun exposure. There are 4 prior reports of basal cell tumors in nonhuman primates; this report is novel in that the tumor is actually confined to an area of the body consistent with the most common form of the disease in humans. In previous reports the animals were also wild caught and actual exposure history and age of the animal were unknown.

Physiological and Behavioral Effects of Social Introduction on Adult Male Rhesus Macaques (*Macaca mula a*)

LA Doyle*

Tulane National Primate Research Center, Covington, LA

Pair housing of laboratory macaques is widely considered to lead to positive changes in wellbeing, yet the process of introduction is viewed as potentially stressful and risk-prone. Behavioral and physiological data were collected on 8 adult male rhesus macaques before, during, and after the process of introduction in order to measure both the initial stress of introduction and long-term changes in well-being. Previously corral-housed subjects, all implanted with biotelemetry devices, were studied in five successive phases: baseline (single housing), 1 d each of protected contact and full contact introduction, post-introduction (1-3 wk after introduction), and settled pairs (≥20 wk after introduction). We coded 176 h of focal animal behavioral data and analyzed 40 h of heart rate data. Fecal cortisol levels were also measured for the baseline, post-introduction and settled-pair phases. All introductions were successful and the process of social introduction showed no physiological or behavioral signs of stress, such as increased heart rate or elevations in depressive, anxiety, or abnormal behaviors. Agonism was minimal throughout the introduction process and over the subsequent months; only 1 wound was incurred over the course of the study. Both short- and long-term improvements in levels of depressive, locomotor, and affiliative behavior were observed. Cortisol levels were significantly lower than baseline in the post-introduction and settled pair phases. Abnormal behavior fell during and after introductions, but this reduction did not persist long-term and may have been confounded by increasing duration of time spent caged. Heart rate varied predictably over the course of each day but not over study phases. The outcome of introductions and long-term improvements in well-being suggest that managers should not be hesitant to introduce adult male rhesus macaques into pairs. The results of this study may be of practical use for designing and monitoring social introductions.

Multiple Congenital Deformities in a Newborn Rhesus Macaque (Macaca mula a)

LD Martin*, L Colgin, C Doane, G Maginnis

Oregon National Primate Research Center, Beaverton, OR

A newborn female rhesus macaque (*Macaca mula a*) was presented from outdoor corral housing the morning following birth with an abnormal appearance and overall ill thrift. Upon examination, a midline defect was noted extending from the umbilicus to the chin of the infant, with only a thin layer of tissue covering the thoracic viscera. The newborn was significantly depressed and displayed labored respiration. Due to the severity of the defect and the grave state of the patient, humane euthanasia was pursued. At necropsy gross lesions included sternoschisis, scoliosis, and a markedly arched palate. While in many human cases of sternal clefts there are concurrent cardiac defects, such as ectopia cordis, no cardiac anomalies were noted in this macaque. Possible defects in embryology leading to these congenital abnormalities will be presented, and treatment options and prognosis in human patients with sternoschisis will be discussed.

Gestational Trophoblastic Disease in an Adult Female Cynomolgus Macaque (Macaca fascicularis)

J Hutt, C Emerson*, J Bayardo

Lovelace Respiratory Research Institute, Albuquerque, NM

An adult female cynomolgus macaque was evaluated for lethargy and dehydration. Results of blood work were unremarkable, and diarrhea was suspected as the cause of the dehydration. The animal's clinical signs improved the day after admission, but declined dramatically by the third day and the animal was euthanized. At necropsy, a sessile uterine mass was identified in the wall of the uterine body, with a portion of the mass extending into the lumen. The mass was approximately 5 cm in diameter, yellowish-white, and friable in the center. Multiple, multifocal coalescing metastatic nodules of similar appearance were detected in the lungs, with effacement of approximately 80% of the normal lung parenchyma. Few metastases were also detected in the liver and spleen. On the basis of histologic appearance of the uterine mass and select immunohistochemical stains, the mass was tentatively identified as a placental site trophoblastic tumor (PSTT). Placental site trophoblastic tumor is one of the gestational trophoblastic diseases. PSTT is a neoplasm of the implantation site commonly following normal pregnancy or abortion. Sex chromosome evaluation suggests that the paternal X chromosome is involved in the development of these tumors.

Compliant Housing of Nonhuman Primates in Preclinical Safety Testing

S Rensing*, W Mueller, A Hartmann

Covance Laboratories GmbH, Muenster, Germany

Although the number of nonhuman primates (NHPs) used for biomedical research is relatively small, the issues which need to be addressed in the discussion of their housing are particularly complex. For toxicology and pharmacology studies, housing and husbandry systems for NHPs should allow the control of as many external variables as possible while facilitating the recording of experimental observations and measurements. The housing should allow easy identification and capture of animals, clear observation and recording of data, and be easy to maintain hygienically. To enhance animal welfare and fulfill morphological, physiological, psychological, and social requirements, a cage design with extended floor area and cage

volume as well as furniture, flexibility, and feeding provisions was designed and optimized through several generations of prototypes in cooperation with manufacturers, technicians, scientists, and veterinarians. In designing such a cage. factors such as sexual dimorphism, age, size, and group dynamics have to be considered. In pharmacological or toxicological investigations, it may be necessary to administer test substances, collect blood samples, and monitor aspects of metabolism over days. The type of procedures and the degree of human intervention involved may affect the type of housing required.

Diagnosis and Treatment of Nephrotic Syndrome in a Male Captive Chimpanzee (Pan troglodytes)

ML Lammey*, DR Lee

Alamogordo Primate Facility, Holloman AFB, NM

Renal disease represents the second leading cause of death in captive chimpanzees and a significant cause of morbidity at Alamogordo Primate Facility (APF). Glomerulonephropathy is the most commonly diagnosed renal abnormality at APF. Nephrotic syndrome is a chronic, progressive clinical sign complex that consists of proteinuria, hypoalbuminemia, hypercholesterolemia, and dependent edema. We present a clinical case in a 32-y-old male chimpanzee (non-infectious), initially observed by animal care staff to have mild bilateral swelling proximal to the tibiotarsal joint. An annual physical examination revealed minimal dependant edema, increased blood cholesterol, and hypoalbuminemia. A subsequent urinalysis showed proteinuria and an increased urine protein to creatinine ratio, demonstrating impaired glomerular filtration. Initially, Lasix (furosemide, 50 mg) was used to control the dependent edema, followed by a treatment regimen of enalapril (10 mg), Lipitor (atorvastatin calcium, 10 mg), and aspirin (243 mg), plus a high-protein, low-sodium diet. Weekly free-catch urine samples are used to monitor the level of proteinuria. We conclude that renal disease, specifically glomerulopathy and nephrotic syndrome, can be medically managed in captive chimpanzee populations.

Fluid-filled Cyst in the Caudal Abdomen of a Rhesus Macaque (Macaca mula a)

JM Bacon*

University of Wisconsin-Madison, Madison, WI

A 17-y-old female, rhesus macaque with a history of intermittent reports of anorexia and an 8-y history of use in reproductive research involving ovarian stimulation and laparoscopic oocyte retrieval presented with an enlarged, irregular, and very firm uterus on abdominal and rectal palpation during a routine exam. Abdominal ultrasound revealed a fluid-filled cyst adjacent to the right ovary. Endometriosis was suspected, but aspiration of the fluid within the cyst resulted in an unexpected diagnosis.

Visceral and Neural Larva Migrans in Rhesus Macaques (Macaca mula a)

AS Gozalo^{1,*}, O Maximova², M St. Claire³, RJ Montali¹, JM Ward¹, WR Elkins⁴, KR Kazacos⁵

¹SoBran, Bethesda, MD; ²Center for Biologics Evaluation and Research, US Food and Drug Administration, Rockville, MD; ³BIOQUAL, Rockville, MD; ⁴Comparative Medicine Branch, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD; ⁵Department of Comparative Pathobiology, Purdue University, West Lafayette, IN

Large ascarid larvae within granulomas were noted histologically in the mesenteric and pancreatic lymph nodes of 13 out of 21 rhesus macaques (Macaca mula a) euthanized as part of an experimental viral vaccine study. In addition, 4 of the 13 monkeys had cerebral granulomas containing nematode larvae similar to those within the lymph nodes. Despite the lesions, the animals did not show clinical signs associated with the parasitic infections. Characteristics of the larvae observed included, on cross-section, a mid-body diameter of approximately 60 to 80 µm, a centrally located and slightly compressed intestine flanked on either side by large triangular excretory columns, and prominent single lateral cuticular alae. The morphology of the larvae was compatible with Baylisascaris species. Baylisascariasis is a well-described infection in animals and humans caused by ingestion of the eggs of the raccoon roundworm, columnaris, is found in skunks and can also cause Baylisascaris procyonis. A similar specie cerebrospinal nematodiasis; however, most reported cases have been due to B procyonis. In our cases the macaques were born free-ranging on an island in the southeastern US where raccoons, but not skunks, were found to be common inhabitants, suggesting that B procyonis was the parasite involved. These cases are examples of low-level or covert Baylisascaris infection as described in humans, and provide further evidence of the existence of this parasite in the southeastern US. Cases of clinical neurologic disease would be possible in this colony if any macaques become infected with higher dosages of eggs.

Husbandry and Veterinary Procedures for a BSL-3 Macaque Facility

L Halliday*, J Purcell, JD Fortman

University of Illinois at Chicago, Chicago, IL

This presentation will review the husbandry and veterinary procedures developed to support a Biosafety Level 3 (BSL-3) facility designed for infectious disease research in macaques. The facility contains 2 animal rooms that together house up to 28 macaques, 2 research laboratories, a necropsy room, and various support areas. Procedures to be discussed include: steps to enter and exit the facility, disinfection of equipment and surfaces, cage cleaning and sanitization schedule, decontamination during and after completion of a study, collection of biological samples, and disinfection during necropsy. The presentation will specifically discuss these topics in relation to a project that involved infection with the monkeypox virus. In addition, a brief overview of the disease course and endpoint criteria will be provided.

Unusual Cases

New Innovations in Primary Housing for Great Apes

P Spurlock^{1,*}, WE Britz, Jr.²

¹Centers for Disease Control—Foothills Campus, Fort Collins, CO; ²Britz & Company, Wheatland, WY

This presentation will detail a number of innovations in a newly designed primary housing cage for chimpanzees and other large great apes. The design resulted from a collaborative effort between members of the Centers For Disease Control and Britz & Company (formerly BH, Inc.). The innovations involve primarily the safety for both the housed animal and the caregivers, and the comfort and environmental enrichment for the caged animal while meeting all the requirements for housing these species. Specifically, these innovations include a modularized construction, a newly designed motor-driven full back panel squeeze mecha-

nism, a different approach to the design of guillotine doors, a secure connection for easily attaching transfer units to the main cage, and enrichment additions for food presentation, resting boards, and the attachment of blood drawing sleeves and manipulata.

Invasive Primate Neuroscience: Is More Regulation or Better Management the Key to Success?

PE Honess*, S Wolfensohn

Oxford University, Oxford, Oxfordshire, UK

The UK has a reputation for having some of the strictest regulations in the world governing the use of nonhuman primates in research. The rationale for this is that the work will then be better justified, carried out to higher standards and result in better science and better animal welfare. However, the primary key to ensuring the research that takes place is strongly justified and ethically sound is a strong institutional management system, which can be an anachronism in some sections of academia. This case study will illustrate many of the challenges associated with providing good quality veterinary care and behavioral management for primates on neuroscience projects. We will show how use of the various levels of communication open to us can achieve long-term benefits in animal welfare and scientific output and a reduction in reputational risk to the institution. We will demonstrate that more regulation is not always better regulation, and that it is the institution's culture of care that makes the difference.

Surgical Placement of Vascular Access Ports in Common Marmosets, Lessons Learned

M Sauer*, H Dulac, S Aborn, J Doherty, M Batchelder

Bristol-Myers Squibb, Wallingford, CT

Common marmosets were chosen as a primate model for pharmacokinetic (PK) studies. To facilitate taking multiple blood samples in these studies, vascular access ports (VAPS) were implanted. Originally only the femoral artery was catheterized, but as the surgery became more routine we placed two VAPS-one in the femoral artery and one in the femoral vein. This is only the second report of successful vascular access port placement in the marmoset. The first report used the VAP solely to administer drugs and, VAPS only remained in place for three weeks. In our study, male marmosets weighing 250-400 grams were implanted with vascular access ports. Port heads made of titanium and polysulfone were placed subcutaneously in the dorsal lumbar area. The VAPS had hydromer coated and heparin coated 3-3.5 French polyurethane catheters attached. The profile of the port head was an important consideration for increasing port longevity. Catheters were locked with a taurolidine-citrate catheter solution during surgery. Catheters are maintained weekly and locked with heparin. Two of the challenges with this model included weight loss and tissue necrosis over the port heads. To minimize weight loss, we provided our marmosets with dietary supplementation that included fortified baby cereal, human nutritional supplement drinks, yogurt, and acacia gum with added protein. To reduce the incidence of tissue necrosis we used a VAP with a lower profile port head. We presently have a colony of 9 ported marmosets. The marmosets have been used successfully twice a month on studies without adverse effects.

CDC's Role in Regulating the Importation and Quarantine of Nonhuman Primates

G Galland*

Centers for Disease Control and Prevention-Atlanta, GA

The importation of nonhuman primates has been controlled by the Centers for Disease Control and Prevention (CDC) since 1975. Regulations were implemented in response to concerns about Ebola-Marburg, monkeypox, yellow fever, and tuberculosis. Following an outbreak of Ebola-Reston in imported cynomolgus macaques in 1989-1990, the requirements to import nonhuman primates were strengthened. This presentation provides an overview of CDC regulations regarding the importation of live nonhuman primates, the scientific rationale for quarantine requirements, and current data on the numbers of registered importers, species and animals imported, and countries of origin. To ensure safe handling of nonhuman primates and compliance with federal quarantine regulations, CDC requires importers to register and provide advance notice of all shipments of animals. As part of becoming a registered importer, CDC requires that each facility submit import plans, disease control measures, standard operating procedures, and facility information for review. In addition, the CDC Division of Global Migration and Quarantine's Zoonoses Team inspects the facilities of each registered importer, monitors shipments of imported animals, and keeps records of numbers and sources of animals imported, as well as morbidity and mortality data. Once registered, an importer must inform CDC about arrival dates, flights, number of animals, ports of entry, ground transport, source, and any illness or death that occurred during transportation or at any time during the quarantine period. Imported animals must be quarantined for a minimum of 31 days, monitored for signs of disease, and treated or tested as necessary. Filovirus infections (including Ebola and Marburg) are primary zoonoses of concern; both have an incubation period of 14 days. Thus, the 31-day guarantine requirement ensures that animals released from quarantine do not pose a risk for transmission of these viral hemorrhagic fevers. Additionally, each animal must have three consecutive negative tuberculin skin tests administered at two-week intervals before release from quarantine. If initiated early, these tests can be completed during the minimum required 31-day period. If any animals test positive for tuberculosis during quarantine, the entire cohort must be held in extended quarantine and undergo additional testing to rule out tuberculosis. Since the CDC regulations were instituted, the health of imported nonhuman primates has improved steadily, and morbidity and mortality rates among them have declined to less than one percent, greatly decreasing the risk to public health. These regulations provide the basis for ensuring that imported nonhuman primates do not pose a zoonotic disease risk.

Diagnostics 101: A Laboratory-users' Guide to Pathogen Testing in Nonhuman Primates

Joe Simmons¹, Nick Lerche², Bob Peters³, Julia Hilliard⁴

¹Charles River Laboratories—Wilmington, MA; ²California National Primate Research Center—Davis, CA; ⁴Georgia State University—Atlanta, GA

For reasons of animal health, occupational safety, and research integrity, the testing of nonhuman primates for a variety of infectious agents has become a mainstay of sound colony management for animals used in biomedical research. Testing for targeted agents often involves indirect methods such as antibody detection, direct methods that detect the actual pathogen or some combination of both. The veterinarian or colony manager is

other presentations

frequently faced with a daunting array of choices when requesting services from a testing laboratory. Equally daunting is the task of translating test results into management decisions. This seminar will provide an overview of some basic concepts underlying laboratory testing of nonhuman primates. Topics will include a discussion of the various types of tests (e.g. screening vs. diagnostic; EIA, IFA, immunoblot, culture, PCR) and the strengths and limitations of each; test characteristics such as sensitivity, specificity, and predictive value; test validation and development of interpretive criteria; and the development of testing algorithms that utilize multiple tests. The objective of this seminar is to provide the veterinarian or colony manager with basic information about diagnostic tests and the testing process that can serve as the basis for improving communication between those charged with utilizing test results and those charged with generating them.

Cardiac Disease in New World Monkeys

AS Gozalo, DVM

¹SoBran, Bethesda, MD

Cardiovascular diseases in New World monkeys are much more prevalent than what has been reported in the literature. Among the neotropical primates, the owl monkey is well-known for developing cardiomyopathy in captivity. Clinically silent cardiovascular pathology is also found in tamarins and squirrel monkeys. It is very likely that most primate species suffer from some degree of cardiovascular disease in captivity. This has been overlooked because most cases are subclinical, and at microscopic examination, in many instances, subtle changes are observed. Evaluation of heart function in monkeys using cardiac enzymatic activity is problematic because although published values exist for CK, AST, and LDH in some primate species-particularly macaques-there are no validated reference values for healthy animals. Most of the values reported have a wide range, and it is difficult to know if these values reflect a healthy population. The sampled animals are described as "clinically healthy," but because they rarely show clinical signs of disease, particularly cardiac disease, they may not be. In addition, there are no published "normal" reference values for CK/AST ratio in nonhuman species, which is useful for the diagnosis of myocardial infarcts in humans, because the CK/AST ratio is not used in veterinary medicine. Although hypertension is suspected as the primary cause of cardiomyopathy in captive monkeys, it is difficult to prove, due to a lack of information on "normal" blood pressure values for nonhuman primates. Echocardiography, electrocardiograms, and X-rays have all been used, but do not clearly identify monkeys with early cardiac disease. In addition, myocardial infarction can occur with normal coronary arteries. The possible mechanisms causing myocardial infarction with normal coronary arteries are hypercoagulable states, coronary embolism, imbalance between oxygen demand and supply, intense sympathetic stimulation, non-atherosclerotic coronary diseases, coronary trauma, coronary vasospasm, coronary thrombosis, and endothelial dysfunction. Cardiac disease in captive nonhuman primates is probably much more prevalent than originally thought, due to the difficulty of making an antemortem diagnosis because of a lack of specific criteria and validated normal reference values.

Nonhuman Primate Breeding in China: the Status and Prospective

WZ Fang

Although China has a long history of breeding nonhuman primates, it has become one of the largest suppliers of NHPs only in recent years. China's colony size has expanded rapidly from 2004, when the central government issued the Provision 124, to enforce the law and to introduce the market mechanism to the industry, setting up a quota system under which a sales quota is issued to every breeding farm according to its colony size and the productivity; thus, encouraging the farms to enlarge its size by acquisition or importation. Many high standard facilities were built, and the total colony size is up to 150,000, which consists of about 120 heads of cynomolgus, about 30 heads of rhesus, and the rest of other species, according to a recent survey conducted by the Chinese Laboratory Primate Association (CLPA), a nongovernmental entity which has over 40 breeding farms and related organizations as it members. CLPA acts as a bridge and liaison between the government and breeding farms, assists government to establish the quota system and supervise the market, and develops academic exchanges among members and end-users. With the favorable investment environment and availability of skilled people, China will supply more and more quality animal and seize the new opportunities in the CRO area.

The Dynamics of Endodontic Treatment

M Alves, DVM

¹UIC College of Dentistry, Chicago, IL

There is no evidence-based protocol for pulp preservation or endodontic procedures for primates. Present literature is not enough to support standards for diagnosis and indication of treatment. What is in use now was adapted from humans. The endodontic specialty had a remarkable technological evolution focused on efficiency and speed of treatment in the past 10 years. This modernization reduced the chair time significantly and radicalized the procedures towards pulp elimination. Modern endodontics does not preserve pulp, but eliminates it. The basic principle of dentistry is still the same-preserve pulp vitality or seal the canal to prevent dead space from where infection can start. Pulp vitality and viability are directly related to tooth long-term survival. Pulpotomy or root canal are sensitive procedures and can fail after minor mistakes. To reduce the chances of mistakes, the adoption of the most accurate diagnostic system and least complicated protocol of treatment should be considered. In the 1980s the mutilating extractions and the failing of pulpal mummification using formocresol on cut canines due to its misuse spurred a search for a permanent solution. Mummification was used, and still is, to treat deciduous teeth with vital pulp exposed during dental procedures as a short-term treatment to prevent infection. The objectives of this presentation are to quickly review the scattered information available, discuss the pros and cons of different techniques, and start a discussion of a dedicated protocol for primate endodontics. The solution is flexibility of treatment with options for temporary (emergency), conservative (vital), permanent, and radical.

Mario Alves Clinical Professor of Periodontics UIC College of Dentistry 801 South Paulina St Chicago, IL 60612 Phone: (312) 996-0745 Fax: (312) 996 -0943 Email: alves@uic.edu Jennifer Bacon Veterinarian Resident

Wisconsin National Primate Research Center 1220 Capital Court Madison, WI 53715 Phone: (608) 209-7162 Email: jbacon@primate.wisc.edu

Kirstin F Barnhart Assistant Professor UT MD Anderson Cancer Center 601 Cool Water Dr Bastrop, TX 78602 Phone: (512) 321-3991 Fax: (512) 332-5397 Email: kfbarnha@mdanderson.org

Bruce J Bernacky Assistant Professor UT MD Anderson Cancer Center Veterinary Sciences 650 Cool Water Dr Bastrop, TX 78602-6621 Phone: (512) 321-3991 Fax: (512) 332-5260 Email: bbernack@mdanderson.org

Rebecca S Blackwood Veterinarian Resident III University of California-Davis Primate Medicine One Shields Ave Davis, CA 95616 Phone: (530) 752-9510 Fax: (530) 752-8201 Email: rsblackwood@primate.ucdavis.edu Alan G Brady Associate Professor University of South Alabama College of Medicine Comparative Medicine University Biological Resources 307 University Blvd MSB 992 Mobile, AL 36688-0001 Phone: (251) 460-6239 Fax: (251) 460-7783 Email: abrady@southal.edu

Jacquie Calnan President Americans for Medical Progress (AMP) 908 King St Alexandria, VA 22314-3067 Phone: (703) 836-9595 Fax: (703) 836-9594 Email: amp@amprogress.org

Saverio V Capuano III Attending Veterinarian/Associate Director Wisconsin National Primate Research Center Animal Services 1220 Capitol Court Madison, WI 53715 Phone: (608) 263-3571 Fax: (608) 265-2067 Email: capuano@primate.wisc.edu

Joyce K Cohen Associate Veterinarian Emory University Yerkes National Primate Research Center 954 Gatewood Rd Atlanta, GA 30329 Phone: (404) 712-8103 Fax: (404) 727-3756 Email: joyce.cohen@emory.edu

Lynn A Collura Lab Animal Medicine Resident Yerkes National Primate Research Center 615 Michael St NE Ste G02 Atlanta, GA 30322 Phone: (404) 727-9057 Fax: (404) 727-3212 Email: lcollur@dar.emory.edu Christina L Cruzen Research Animal Veterinarian Wisconsin National Primate Research Center Veterinary Services 1220 Capitol Court Madison, WI 53715 Phone: (608) 265-0622 Fax: (608) 265-2067 Email: ccruzen@primate.wisc.edu

Lara A Doyle Veterinarian Resident Tulane National Primate Research Center 18703 Three Rivers Rd Covington, LA 70433 Phone: (985) 871-6374 Fax: (985) 871-6328 Email: Idoyle@tulane.edu

Carol L Emerson Veterinarian/Associate Scientist Lovelace Respiratory Research Institute 2425 Ridgecrest St SE Albuquerque, NM 87108 Phone: (505) 548-9611 Fax: (505) 548-4980 Email: cemerson@lrri.org

Weizhen Fang General Secretary Beijing Puliyuan Trading LTD Room 132 E 11 Bldg 83 Fuxing Rd Beijing100039 China Phone: 86(105)1607408 Fax: 86(105)1607403 Email: wzfang2008@yahoo.com.cn

Gale Galland Veterinary Officer CDC 1600 Clifton Rd Atlanta, GA 30333 Phone: (404) 639-4528 Fax: (404) 498-1633 Email: ggg0@cdc.gov

workshop**participants**

Melaney K Gee Assistant Director Wake Forest University Health Sciences Animal Resources Program Medical Center Blvd Winston Salem, NC 27157-0001 Phone: (336) 716-1535 Fax: (336) 716-1501 Email: mgee@wfubmc.edu

Sylvia I Gografe Director Vanderbilt University Office of the Animal Welfare Assurance 1161 21th Ave S, S/1316MCN Nashville, TN 37232 Phone: (615) 322-5499 Fax: (615) 936-8160 Email: sylvia.gografe@vanderbilt.edu

Alfonso S Gozalo Attending Veterinarian SoBran NIAID/CMB 9000 Rockville Pike Building 14BS Rm 228 Bethesda, MD 20892 Phone: (301) 496-9333 Fax: (301) 480-2377 Email: gozaloa@niaid.nih.gov

Rebecca L Grant Associate Director NPRP University of Pennsylvania 125 S 31 St Ste 2018 Philadelphia, PA 19104-3403 Phone: (215) 573-6822 Fax: (215) 573-5655 Email: rgrant2@mail.med.upenn.edu

Lisa C Halliday Clinical Veterinarian University of Illinois-Chicago Biologic Resources Laboratory 1840 W Taylor St MC 533 Chicago, IL 60612-7348 Phone: (312) 996-9453 Fax: (312) 996-8065 Email: Ihall@uic.edu

Julia K Hilliard Professor Georgia State University Virology/Immunology PO Box 4118 Atlanta, GA 30302-4118 Phone: (404) 651-0811 Fax: (404) 651-0821 Email: jhilliard@gsu.edu

Paul Honess Primatologist University of Oxford Department of Veterinary Services Kesselfeld 29 Parks Rd Oxford Oxfordshire OX1 3PT United Kingdom Phone: 44(186)5272545 Fax: 44(186)5272118 Email: paul.honess@vet.ox.ac.uk

Kelly M.S. Hugunin Postdoctoral Scholar University of Michigan ULAM 1150 W Medical Center 018 ARF Ann Arbor, MI 48109 Phone: (734) 936-1696 Fax: (734) 936-3235 Email: kellyhug@umich.edu

Michael L Lammey Clinical Veterinarian Charles River Laboratories Alamagordo Primate Facility PO Box 956 Building 1303 Holloman AFB, NM 88330 Phone: (505) 679-3800 Fax: (505) 679-3841 Email: mlammey@criver.com

D Rick Lee Director Alamogordo Primate Facility PO Box 956 Holloman AFB, NM 88330-0956 Phone: (505) 679-3801 Fax: (505) 679-3841 Email: rick.lee@crl.com Nicholas W Lerche Associate Director University of California-Davis California National Primate Research Center 855 Malaga Ave Davis, CA 95616-8542 Phone: (530) 752-6490 Fax: (530) 752-4816 Email: nwlerche@ucdavis.edu

Gwendalyn M Maginnis Chief Clinical Medicine Unit OHSU/Oregon National Primate Research Center 505 NW 185th Ave Beaverton, OR 97006-3448 Phone: (503) 690-5221 Fax: (503) 690-5264 Email: maginnis@ohsu.edu

Keith Mansfield Associate Professor Pathology New England Primate Research Center 1 Pine Hill Dr Southborough, MA 01772 Phone: (508) 624-8183 Fax: (508) 624-8135 Email: keith_mansfield@hms.harvard.edu

Lauren Drew Martin Veterinary Intern Oregon National Primate Research Center 505 NW 185th Ave Beaverton, OR 97006 Phone: (503) 939-5561 Fax: (503) 614-3736 Email: martilau@ohsu.edu

Preston A Marx Tulane Regional Primate Research Center 18703 Three Rivers Rd Covington, LA 70433 Phone: (985) 871-6255 Fax: (985) 871-6248 Email: pamarx@tpc.tulane.edu

Margaret S McTighe Clinical Veterinarian Vanderbilt University Division of Animal Care 1161st 21st Ave S AA-6206 MCN Nashville, TN 37232-0001 Phone: (615) 322-7255 Fax: (615) 343-7682 Email: maggie.mctighe@vanderbilt.edu

Nancy L Merrill Madigan Army Medical Center Clinical Investigation 9040 Fitzsimmons Tacoma, WA 98431 Phone: (253) 968-0145 Fax: (253) 968-1044 Email: nancy.l.merrill@amedd.army.mil

Lorna Millen Veterinarian Covance Research Products Jim Wells County Rd #381 Alice, TX 78333 Phone: (361) 664-4985 Fax: (361) 664-8933 Email: lorna.millen@covance.com

Pablo R Morales Associate Director Mannheimer Foundation Veterinary Services 20255 SW 360 St Homestead, FL 33034 Phone: (305) 245-1551 Fax: (305) 245-9423 Email: moralesveterinarian@bellsouth.net

Joseph T Newsome Clinical Director University of Pittsburgh Laboratory Animal Resources 1040 S Biomedical Science Tower 3500 Terrance St Pittsburgh, PA 15261 Phone: (412) 648-8950 Fax: (412) 648-8441 Email: newsomej@dlar.pitt.edu Thomas E Nolan Vice President Instech Solomon 5209 Militia Hill Rd Plymouth Meeting, PA 19462-1216 Phone: (610) 941-0132 ext 116 Fax: (610) 941-0134 Email: tnolan@instechlabs.com

Tara G Ooms Clinical Veterinarian Tulane National Primate Research Center 18703 Three Rivers Rd Covington, LA 70433 Phone: (985) 871-6496 Fax: (985) 871-6328 Email: tooms@tulane.edu

Robert L Peters Director Laboratory Animal Health Services BioReliance Corporation 14920 Broschart Rd Rockville, MD 20850-3349 Phone: (301) 610-2225 Fax: (301) 838-0371 Email: bpeters@bioreliance.com

Susanne Rensing Clinical Veterinarian Covance Laboratories GmbH Animal Health Kesselfeld 29 48163 Muenster Germany Phone: (49) 251-97980 Fax: (49) 251-784697 Email: susanne.rensing@covance.com

William C Satterfield
Associate Professor Veterinarian Medical & Surgery
Michale E Keeling Center
Comparative Medicine & Research
650 Cool Water Dr
Bastrop, TX 78602-6621
Phone: (512) 321-3991
Fax: (512) 332-5218
Email: wsatterf@mdanderson.org

Barthel Schmelting Clinical Veterinarian Covance Laboratories KesselFeld 29 48163 Muenster Germany Phone: (49) 251-97980 Fax: (49) 251-784697 Email: barthel.schmelting@covance.com

Steven T Shipley Chief Veterinarian Surgical Services University of Maryland School of Medicine Veterinarian Resources 10 S Pine St MSTF G100 Baltimore, MD 21201-1116 Phone: (410) 706-3703 Fax: (410) 706-8538 Email: sshipley@veterinarianmed. umaryland.edu

Joe H Simmons Director Charles River Lab/Wilmington/Institute Diagnostic Laboratories 251 Ballardvale St Wilmington, MA 01887 Phone: (978) 658-6000 EXT 1776 Fax: (978) 988-9093 Email: joe.simmons@crl.com

Morgan L Singletary Clinical Veterinarian Tulane National Primate Research Center Veterinary Medicine 18703 Three Rivers Rd Covington, LA 70433 Phone: (985) 871-6330 Fax: (985) 871-6328 Email: saicklen@tulane.edu

J Paul Spurlock Team Leader & Attending Veterinarian Centers for Disease Control Animal Resources Branch CDC Foothills Campus 3150 Rampart Rd MS P02 Fort Collins, CO 80521 Phone: (970) 266-3528 Fax: (970) 266-3599 Email: jus2@cdc.gov Stephanie Torreilles Post-Doctoral Fellow Stanford University School of Medicine Comparative Medicine RAF 1 Quad 7 Building 330 Stanford, CA 94305-5410 Phone: (650) 724-4415 Fax: (650) 725-0940 Email: Streetorreil@Streetanford.edu

Brigid V Troan NC Zoological Park Hanes Medical Center 4401 Zoo Pkwy Asheboro, NC 27203 Phone: (919) 522-1933 Fax: (919) 882-9036 Email: brigid@troan.org

Patricia V Turner Ontario Veterinary College Department of Pathobiology University of Guelph Guelph, ON N1G2W1 Canada Phone: (519) 824-4120 Ext. 54497 Fax: (519) 824-5930 Email: pvturner@uoguelph.ca

Joseph L Wagner Director The Mannheimer Foundation Inc Veterinarian Resources 20255 SW 360th St Homestead, FL 33034-4102 Phone: (305) 245-1551 Fax: (305) 245-7650 Email: wagnerj@bellsouth.net

Ruth Ann Woodward Facility Veterinarian NIH NICHD RAMB 2181 Gapland Rd Jefferson, MD 21755-8901 Phone: (301) 496-3750 Fax: (301) 480-3793 Email: woodwarr@mail.nih.gov

Kelly P Yamada Postdoctoral Fellow Memorial Sloan-Kettering Cancer Center 1275 York Ave Box 270 New York, NY 10021-6007 Phone: (646) 888-2417 Fax: (646) 422-0139 Email: yamadak@mskcc.org Maria T Zayas Staff Veterinarian Wake Forest University School of Medicine ARP Medical Center Blvd Winston Salem, NC 27157 Phone: (336) 713-7398 Fax: (336) 713-7395 Email: mzayas@wfubmc.edu



Sth Annual Workshop of the Association of Primate Veterinarians (APV), Oct. 11–13, 2007

This form is provided to attendees of the annual APV workshop as official documentation of attendance at the workshop. Attendees are advised to record their attendance at presentations and to supplement this information with a copy of the official program.

Every hour of participation in the seminars and scientific sessions of an APV workshop is potentially worth one hour of credit for documentation of continuing education. The 2007 workshop provides a total of **16** contact hours of continuing education. Additional information on the various state requirements of CE credits is available from the American Veterinary Medical Association (AVMA). Final determination of the acceptance of a program is up to the individual state licensing authority.

Marion Ratteree, Secretary

Print name of participant Signature of participant apriano TT are of contact person/faculty

At the 2007 APV Workshop, I attended the following lectures and/or presentations:

- Effects of the Macrolide Drug Tylosin on Chronic Diarrhea in Rhesus Macaques, Rebecca S. BlackwoodP
- Treatment of Conjuctivitis by Subtenon Injection
- Using Fenbendazole in Commercial Non-Human Primate Diets and a Fecal Parasite Concentrator to Manage Intestinal Parasites in Macaques
- Asymptomatic Ependymal Cyst in Adult Cynomolgus Macaque
- Trichobezoar Prevention Using Soy Lecithin Supplementation in Galagos
- □ Urinary Alkalinization in the Treatment of Traumatic Rhabdomyolysis in Rhesus Macaques
- □ Localized Allergic Dermatitis Treated with Tacrolimus Ointment in a Rhesus Macaque
- □ Acute Anorexia and Lethargy Associated with Pyrexia in an Adult Rhesus Macaque
- Fractures Associated with a Breeding Colony of SPF Cynomolgus Monkeys
- □ Stress Cardiomyopthy Following Severe Soft Tissue Trauma in a Chimpanzee
- □ Calcinosis Cutis in Rhesus Macaques Associated with Trauma
- Adenocarcinoma in a Chimpanzee
- Management of Iatrogenic Hypoglycemic Coma in a Rhesus Macaque
- Psychosocial Stress and Diet Choice
- □ Acute Renal Failure Associated with Sepsis in Three Rhesus Macaques
- Diagonstic 101: A Laboratory Users' Guide to Pathogen Testing in Nonhuman Primates
- Round-table Discussion: Clinical
- Round-table Discussion: Management
- Techniques for Treating Vaginal Prolapses
- Determining Efficacy of Post-endoscopic Biopsy Treatment with Sucralfate on Appetite in Nonhuman Primates
- Non-Invasive Blood Pressure Determination for Continuous Anaesthesia Monitoring in Nonhuman Primates
- Molecular Epidermicological Investigation of Simian Varicella Virus Infection in Two Immunocompromised Rhesus Macaques
- Some Advances from the Use of Macaques in Gene Therapy

- Limitations of Measles Diagnostic Testing in Rhesus Macaques
- □ Colonoscopy in Rhesus Macaques—Principles, Procedures, and Problems,
- Herpes B Update
- Persistent Conjunctivitis in a Rhesus Macaque
- Focal Pyogranulomatous Encephalitis in a Rhesus Macaque
- Failure to Thrive in a Cynomolgus Monkey
- Abdominal Mass in a Squirrel Monkey
- Round-table Discussion: Cardiac Disease in New World Monkeys
- □ Round-table Discussion: Cardiomyopathy in Captive Chimpanzees
- Irregular Cutaneous Scalp Mass in a Rhesus Macaque
- Physiological and Behavioral Effects of Social Introduction on Adult Male Rhesus Macaques
- Multiple Congenital Deformities Including Sternoschisis and Scoliosis in a Newborn Rhesus Macaque
- Gestational Trophoblastic Disease in an Adult Female Cynomolgus Macaque
- □ Compliant Housing of Nonhuman Primates in Preclinical Safety Testing
- Diagnosis and Treatment of Nephrotic Syndrome in a Male Captive Chimpanzee
- □ Fluid-Filled Cyst in the Caudal Abdomen of a Rhesus Macaque
- Visceral and Nerual Larva Migrans in Rhesus Macques
- □ Husbandry and Veterinary Procedures for a BSL-3 Macaque Facility
- The Dynamics of Endodonic Treatment
- New Innovations in Primary Housing for Great Apes
- □ Invasive Primate Neuroscience: Is More Regulation or Better Management the Key to Success?
- Surgical Placement of Vascular Access Ports in Common Marmosets
- NHP Breeding in China: the Status and Prospective
- AMP Update
- Journal of Medical Primatology: Proceedings and Publications
- ILAR's International Primate Plan

Application is pending to have this program approved by the American Association of Veterinary State Boards (AAVSB) RACE Program for Continuing Education. Please contact AAVSB's RACE Program at 3100 Main Street, Suite 208, Kansas City, MO, 64111 or info@aavsb.org should you have any comments or suggestions regarding this program.